



# ROLE OF MACROPHAGES & LYMPHOCYTES IN RENAL INJURY

By

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Mansoura University

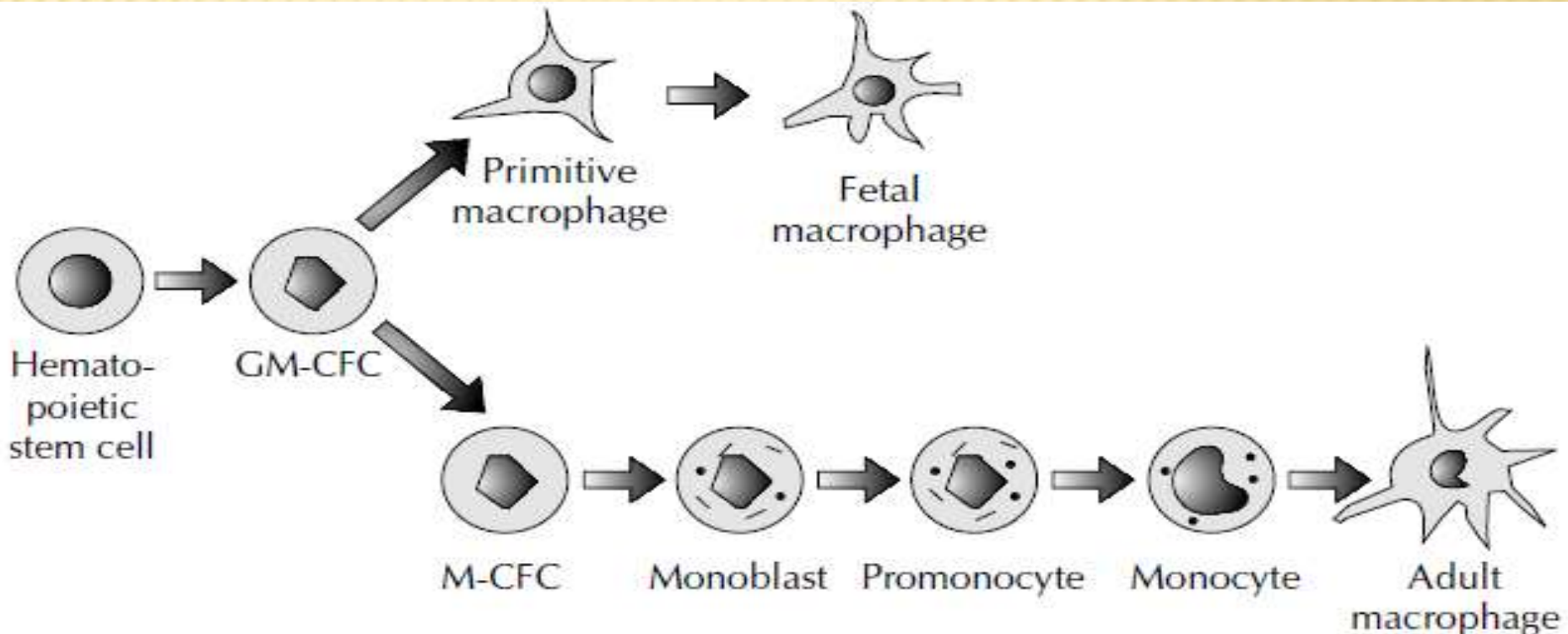
# PHASES OF KIDNEY DISEASES

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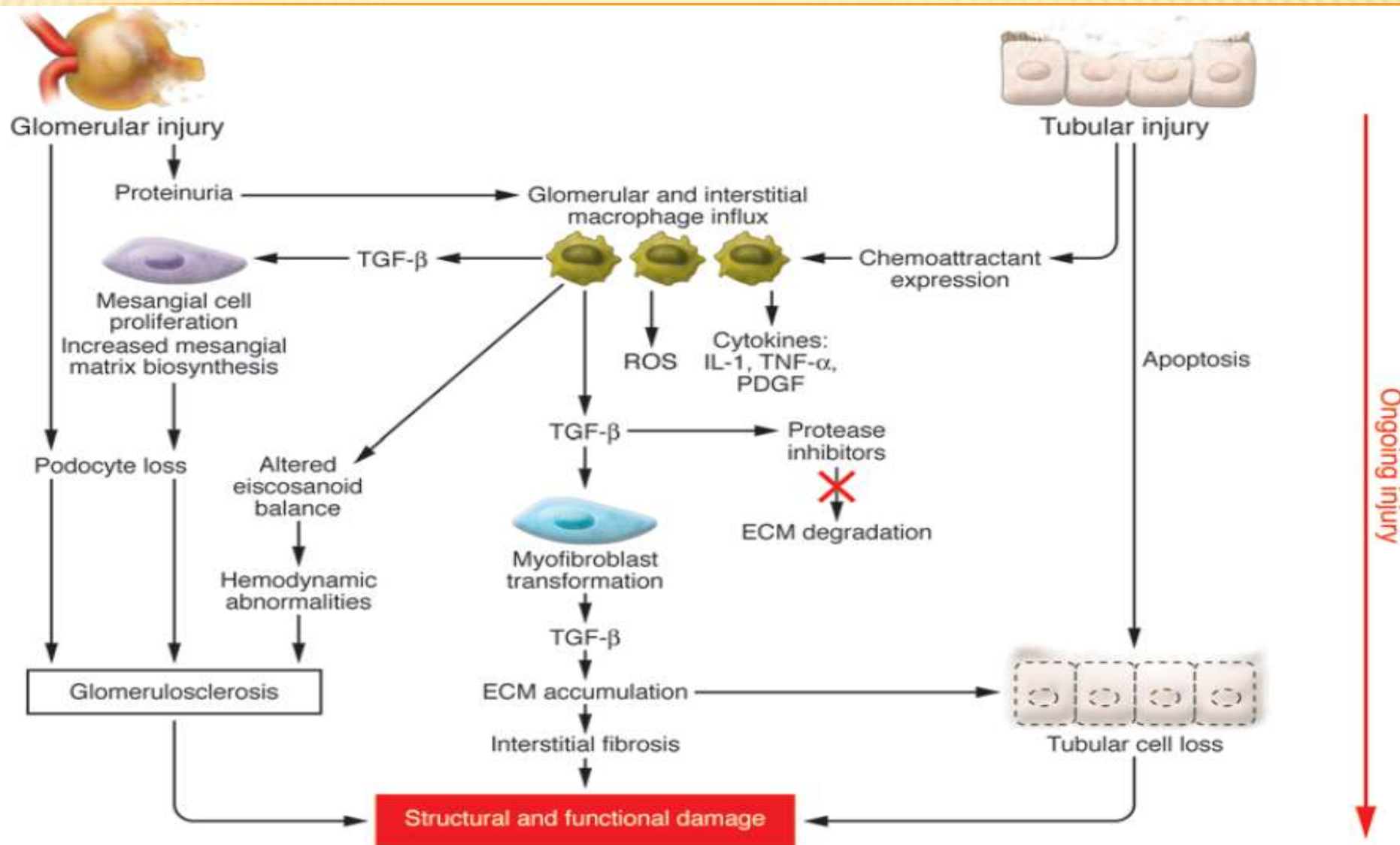
- ✖ Injury phase : tissue inflammation
- ✖ Reparative phase: development of fibrosis
- ✖ Regenerative Phase: proliferation of new cells, derived from either resident cells and/or bone marrow-derived (BMD) cells.

# MACROPHAGES

- ✖ Diverse and dynamic population of cells
- ✖ Perform a wide range of critical functions
- ✖ Secrete a wide range of inflammatory factors.



# ROLE OF MACROPHAGES IN RENAL INJURY





*Cardiovascular, Pulmonary and Renal Pathology*

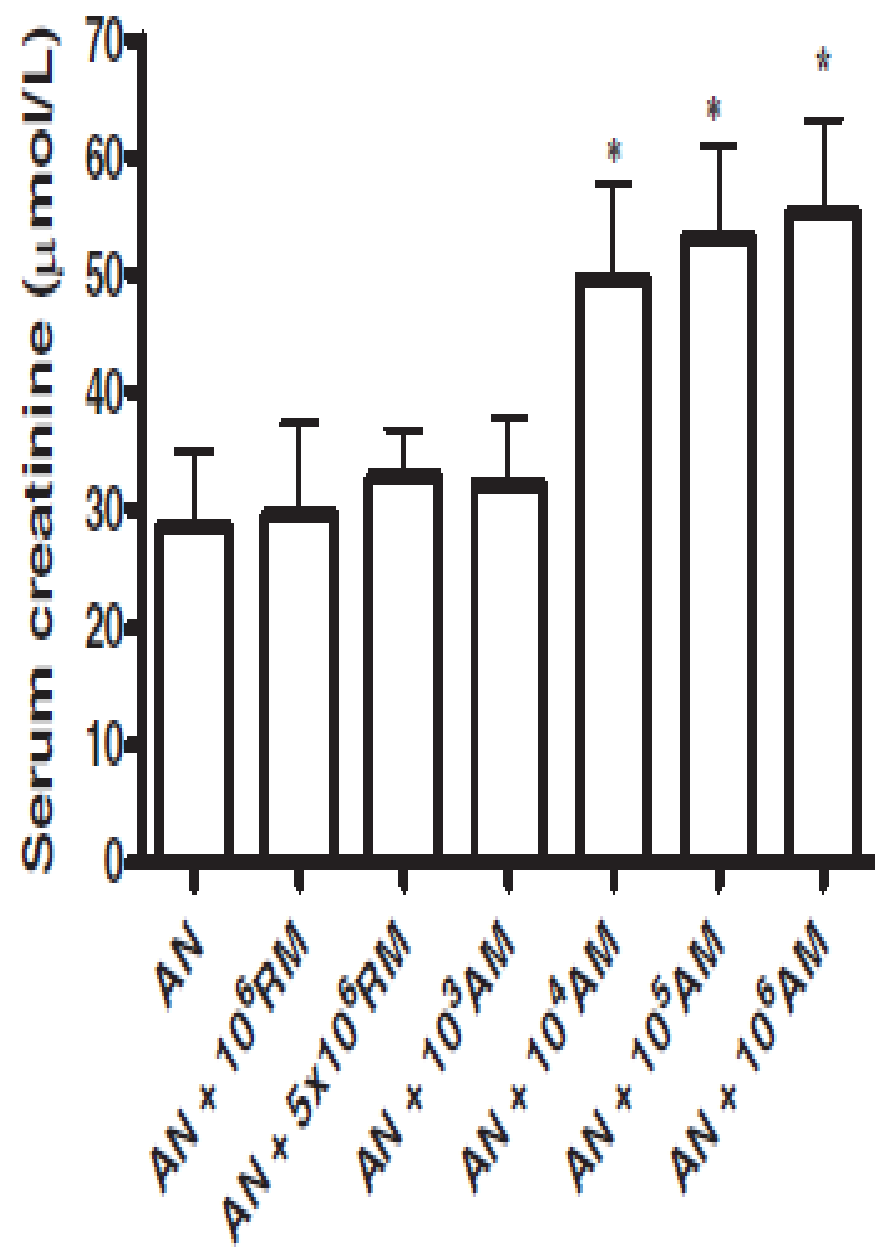
# By Homing to the Kidney, Activated Macrophages Potently Exacerbate Renal Injury

Ying Wang,\* Yiping Wang,\* Qi Cai,\*  
Guoping Zheng,\* Vincent W.S. Lee,\*  
Dong Zheng,\* Xiaomei Li,<sup>†</sup> Thian Kui Tan,\*  
and David C.H. Harris\*

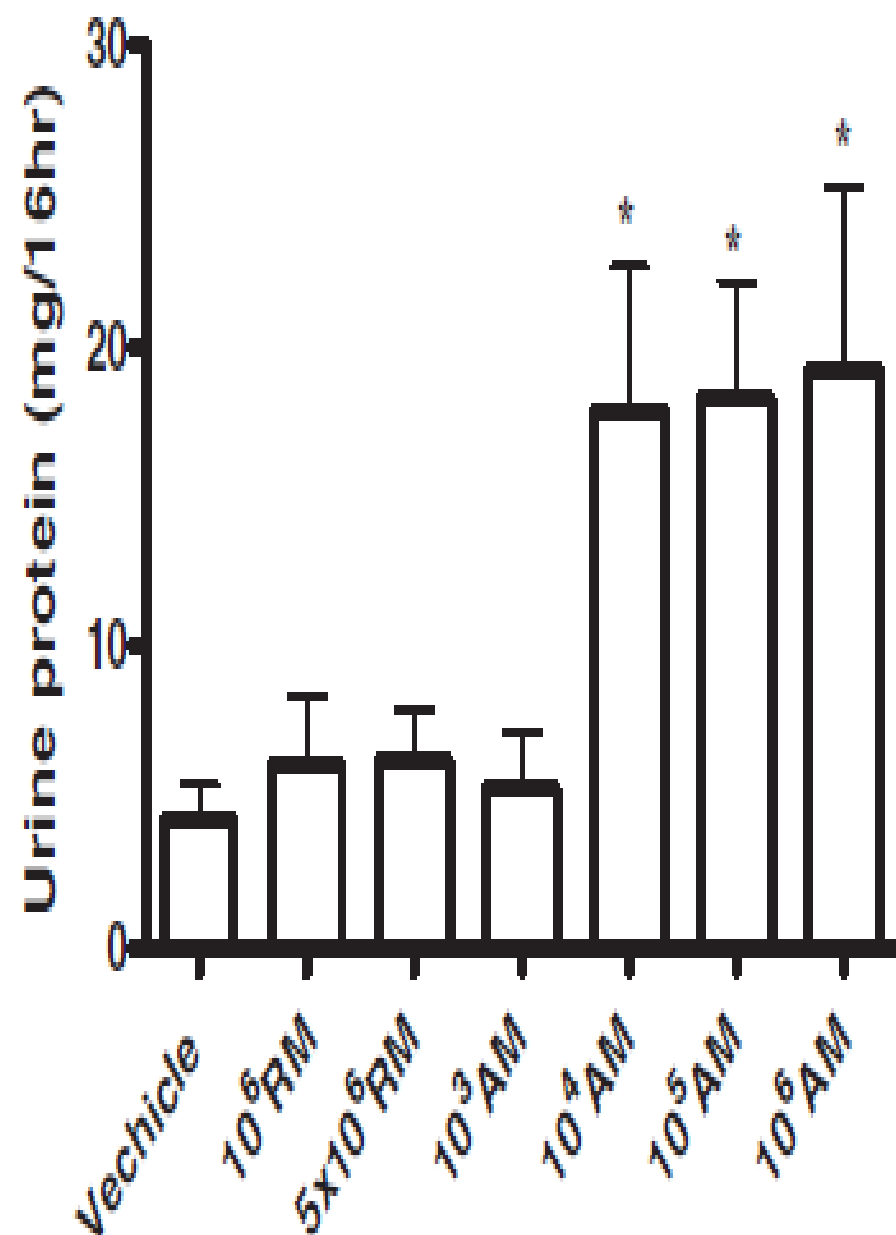
*From the Centre for Transplantation and Renal Research,\*  
The University of Sydney at Westmead Millennium Institute,  
Westmead, Sydney, Australia; and the Department of  
Rheumatology and Immunology,<sup>†</sup> Anhui Medical University at  
Provincial Hospital, Hefei, Anhui, People's Republic of China*

**SCID mice with adriamycin nephrosis, an experimental model of human FSGS were treated intravenously with either resting ( $1 \times 10^6$  to  $5 \times 10^6$ ) or activated ( $1 \times 10^3$  to  $1 \times 10^6$ ) macrophages on day 6 post adriamycin administration, and the effects on kidney injury were examined after 28 days.**

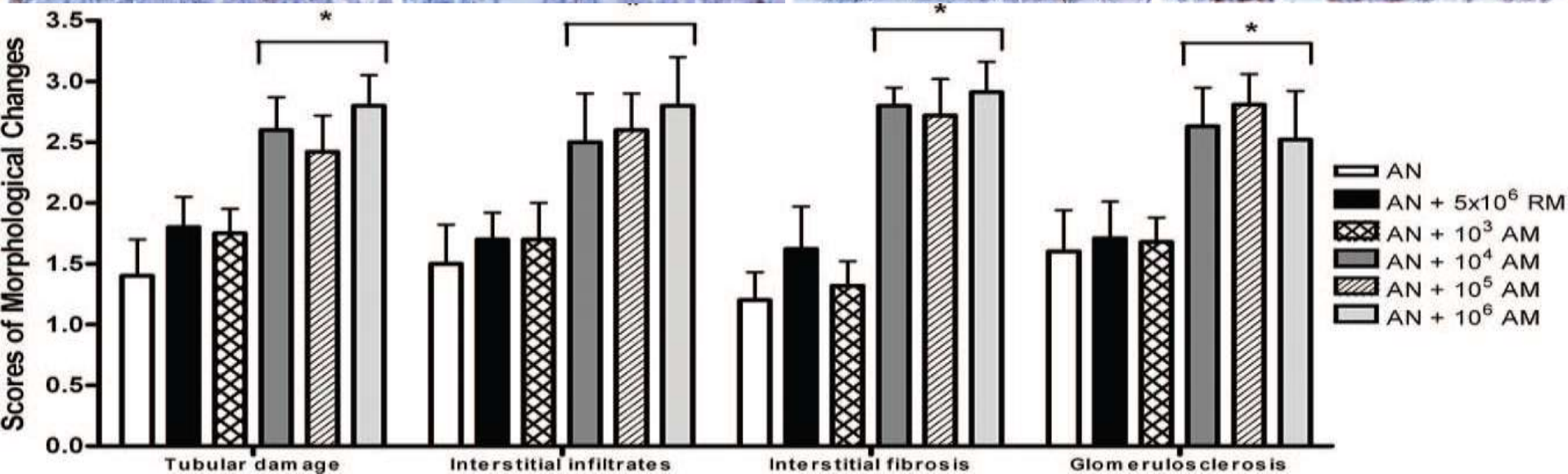
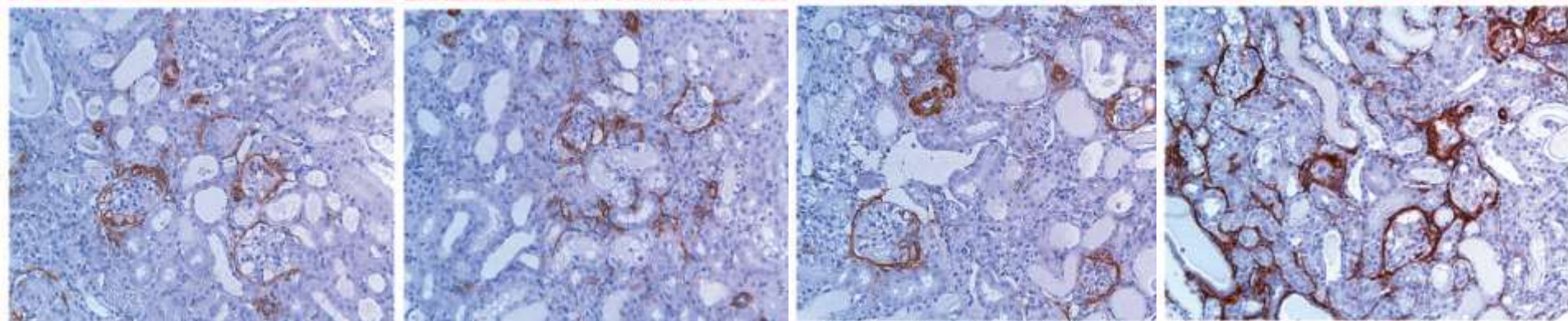
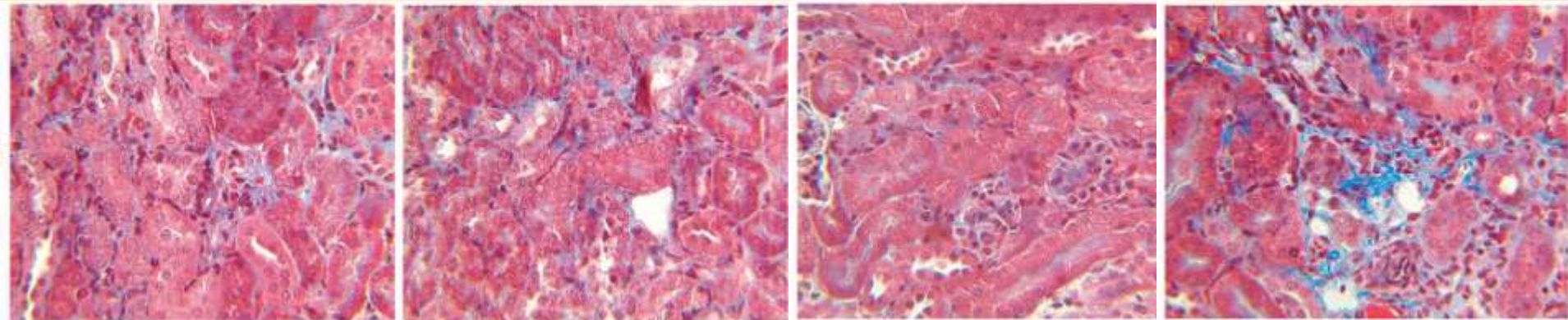
# Serum creatinine



# Urine protein



AN + vehicle

AN +  $5 \times 10^6$  RMAN +  $10^3$  AMAN +  $10^4$  AM



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**DOES MACROPHAGES HAVE ANY ROLE  
IN RENAL REPAIR ??????????**





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Biochemical and Biophysical Research Communications 332 (2005) 11–16

BBRC

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## Adoptive transfer of macrophages ameliorates renal fibrosis in mice

Masashi Nishida \*, Yasuko Okumura, Shin-ichiro Fujimoto, Isao Shiraishi,  
Toshiyuki Itoi, Kenji Hamaoka

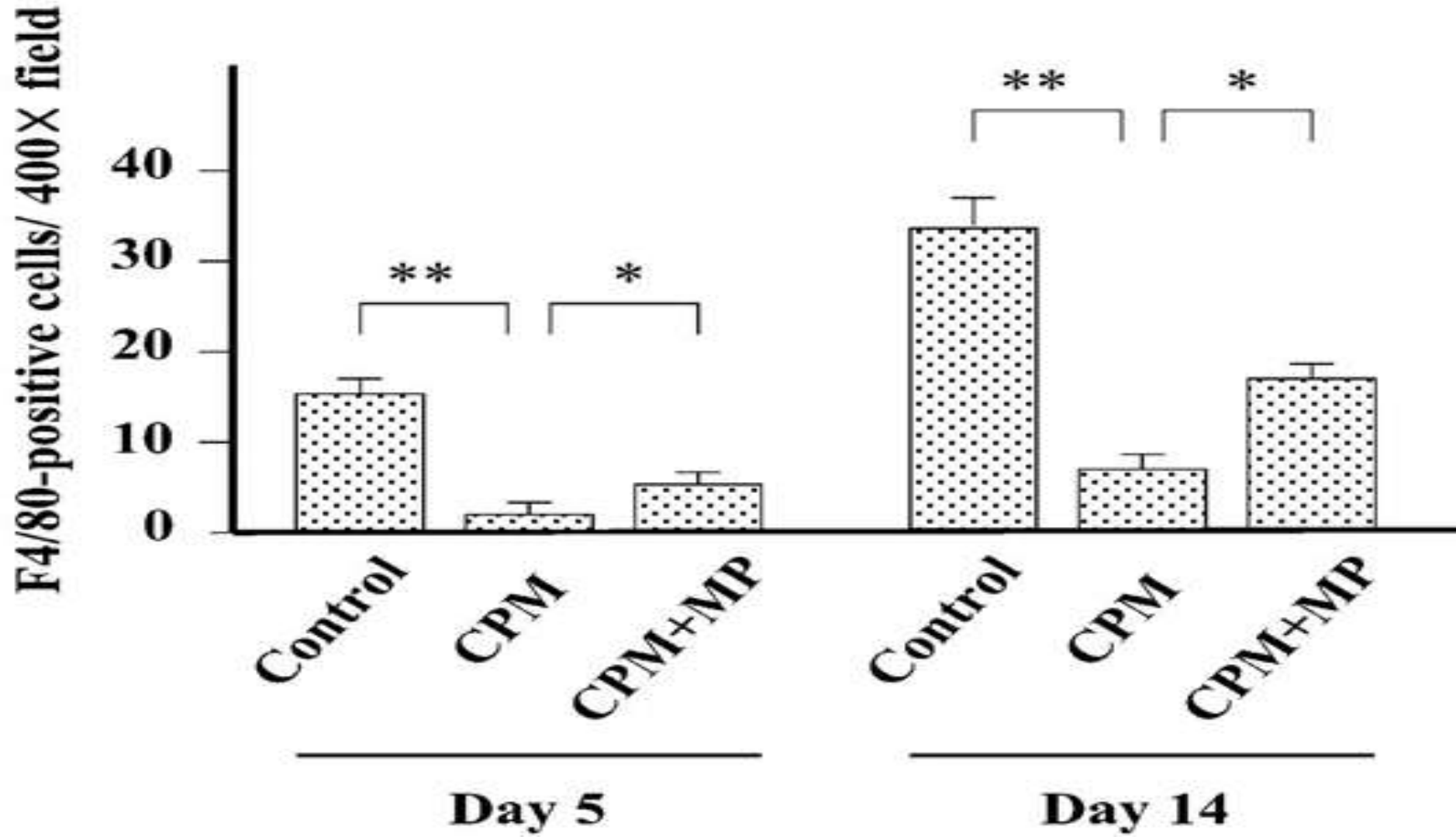
*Department of Pediatric Cardiology and Nephrology, Kyoto Prefectural University of Medicine Graduate School of Medical Science, Kyoto, Japan*

Received 12 April 2005

Available online 26 April 2005

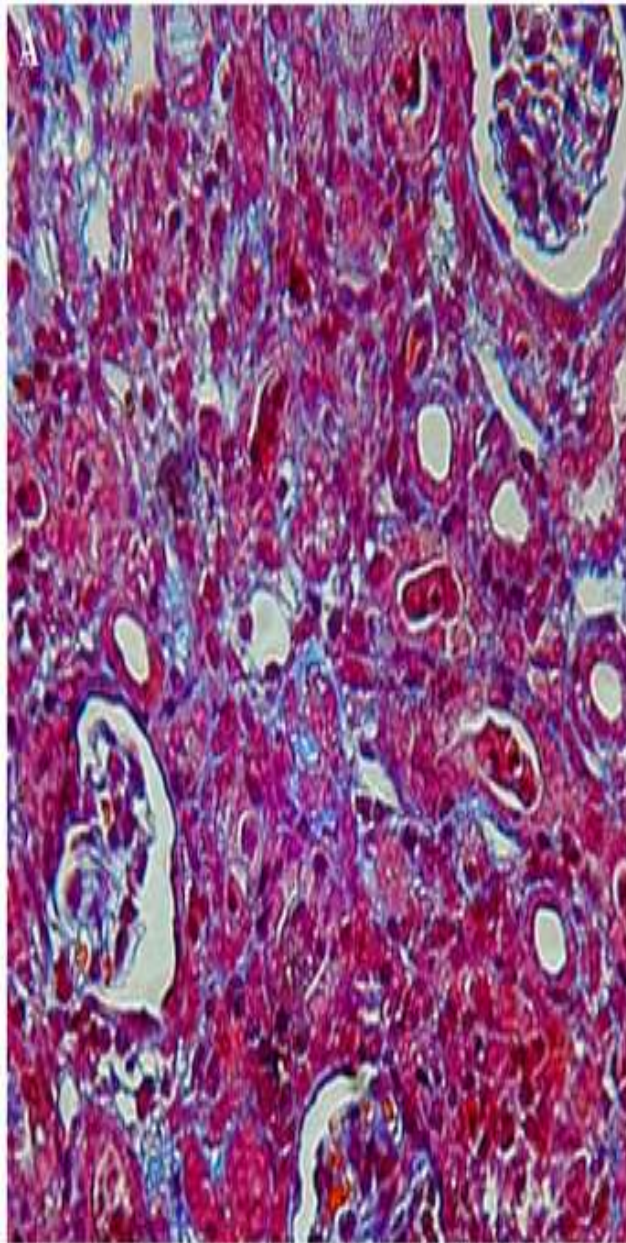
**Adoptive transfer of bone marrow-derived (BM) macrophages following pharmacological depletion of leukocytes by CPM in a C57BL/6 mice model of unilateral ureteral obstruction (UUO) and examination at 5, 14 days.**

# MACROPHAGES INFILTRATING TO THE INTERSTITIUM

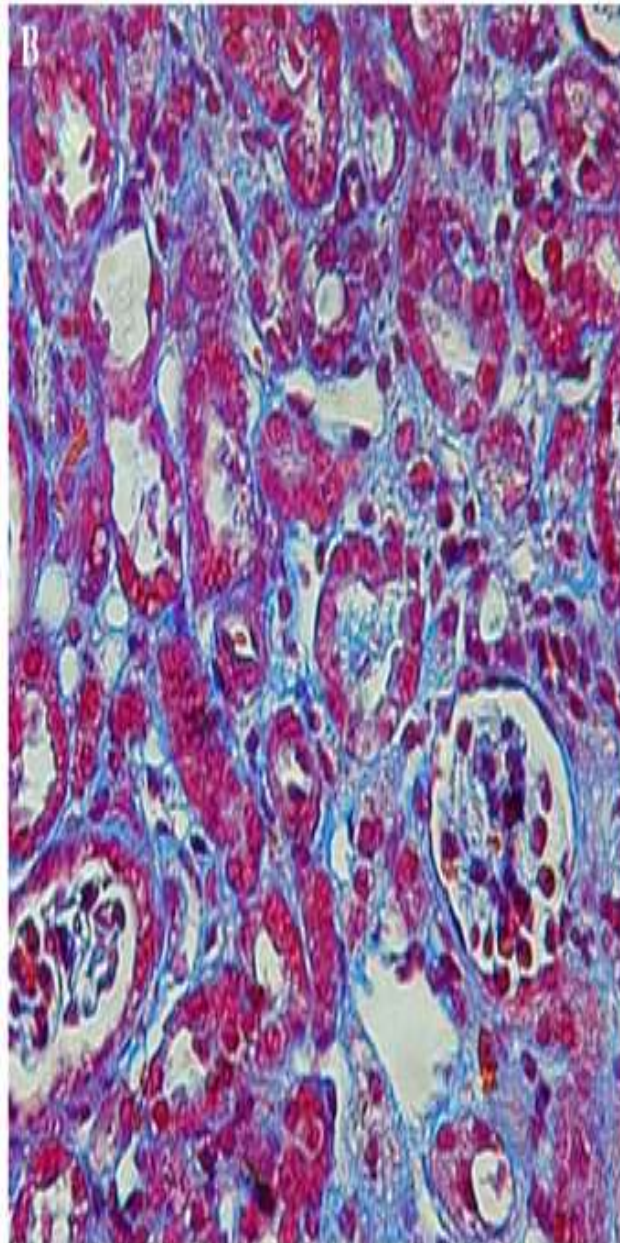




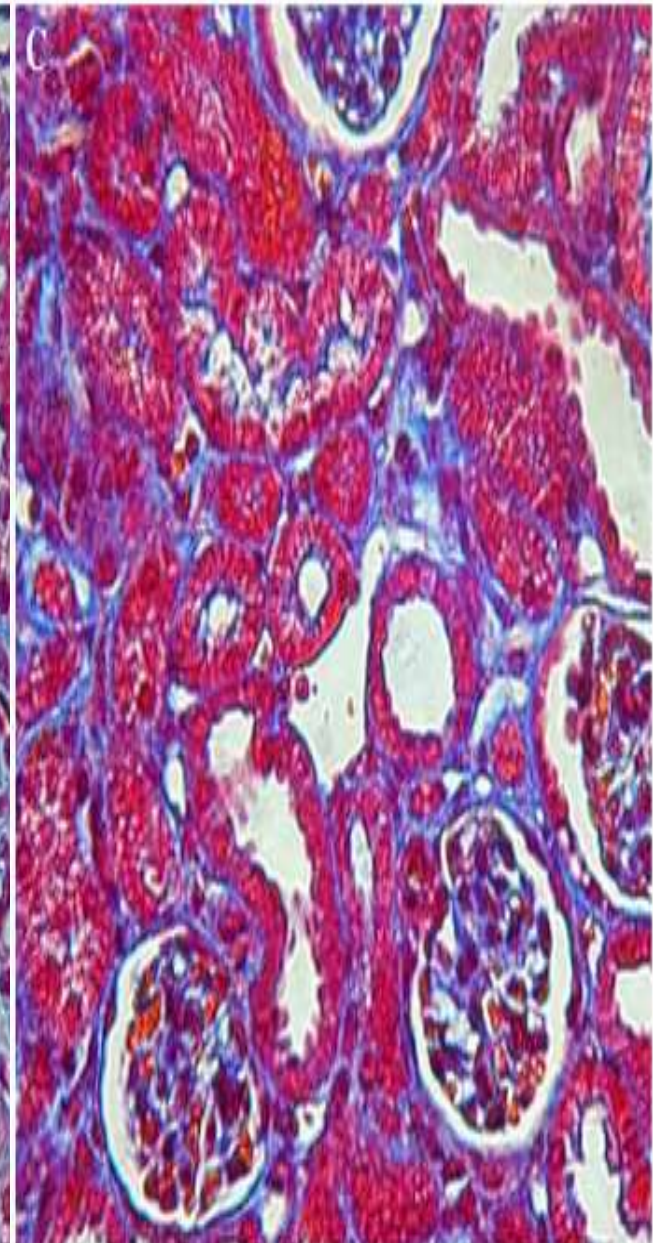
**Control**



**CPM**

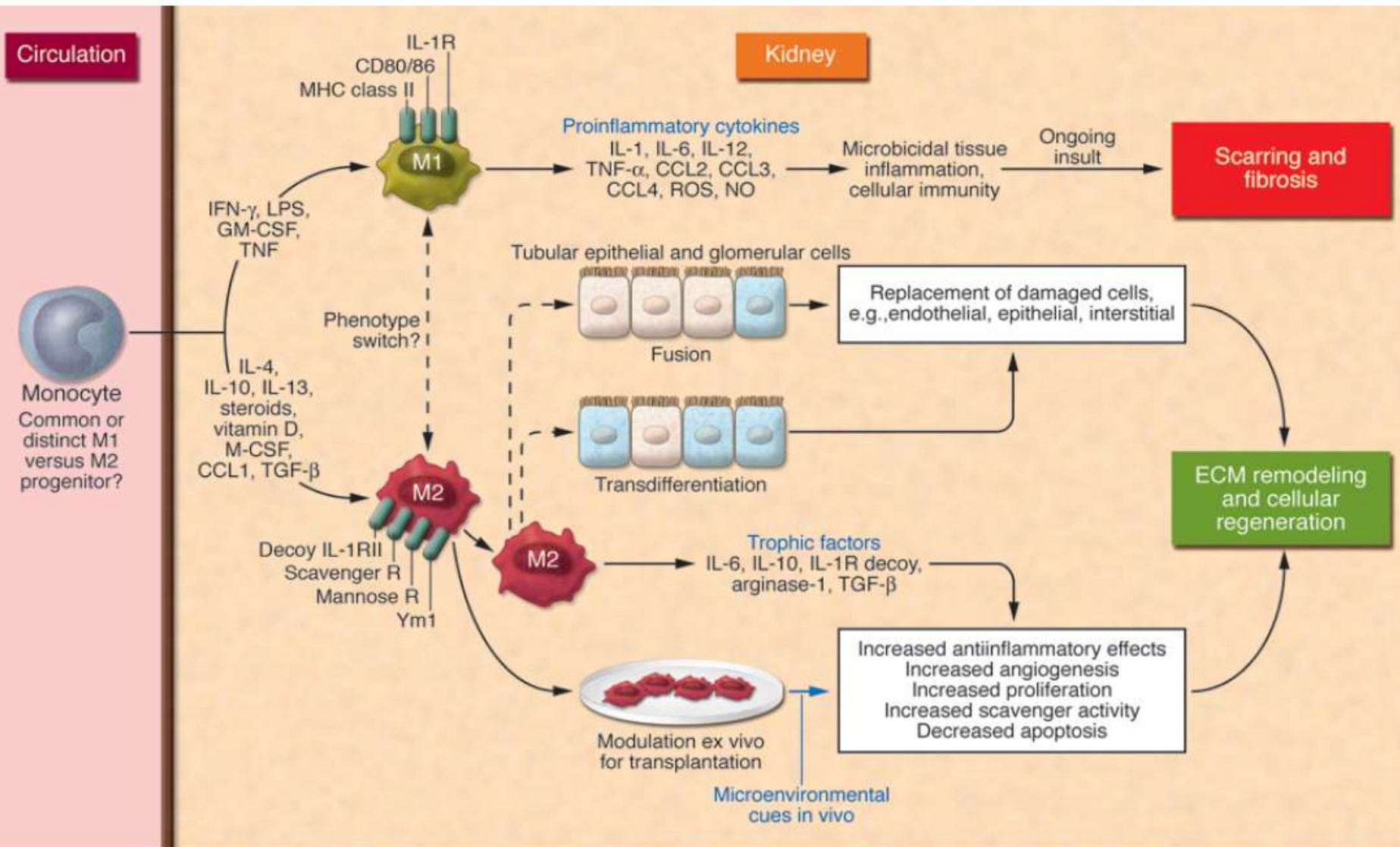


**CPM + MP**





# ROLE OF MACROPHAGES IN RENAL REPAIR





# MACROPHAGE PHENOTYPE

## Macrophage activation states and functions

Macrophage phenotype	Activation state	Stimuli	Phenotypic function	Cytokine and inflammatory profile	Unique surface markers
M1	Classical activation	IFN- $\gamma$ + LPS, TNF, GM-CSF, TLR/IL-1R ligand	Proinflammatory Th1 response	IL-1, IL-12, IL-15, IL-18, IL-23, TNF- $\alpha$ , IL-6, MCP-1, CCL2, CCL3, CCL4, CCL20/MIP-3a, ROS, NO, iNOS, NOS2	CD86, CD80, MHC class II <sup>hi</sup> , IL-1R, IL-12 <sup>hi</sup> , IL-23 <sup>hi</sup> , IL-10 <sup>lo</sup>
M2	Alternate activation (M2a polarization)	IL-4 or IL-13	Th2 responses, type II inflammation	Fibronectin, BIG-H3, arginase-1, TNF- $\alpha$ , IL-6, IGF, CCL13/MCP-4, CCL22, CCL18, $\beta$ 2 integrins	Mannose receptor, scavenger receptor, MHC class II <sup>hi</sup> , decoy IL-1R11, FIZZ1/Ym-1
	Type II activation (M2b polarization)	Immune complex + TLR/IL-1R ligands	Immunoregulation, Th2 activation	IL-10, TNF- $\alpha$ , IL-1, IL-6, IL-12, SPHK1, CCL1	CD86, MHC class II <sup>hi</sup> , IL-10 <sup>hi</sup> , IL-12 <sup>lo</sup>
	Deactivated (M2c polarization)	IL-10, TGF- $\beta$ , glucocorticoids	Immunosuppression, matrix remodeling, tissue repair	IL-10, IL-1 $\beta$ , IL-6, TGF- $\beta$ , ECM proteins, CCL16, CCL18, arginase-1	SLAM (CD150), mannose receptor, MHC class II <sup>lo</sup>

BIG-H3, fasciclin domain 4 protein; MIP, macrophage inflammatory protein; SLAM, signaling lymphocytic activation molecule; SPHK1, sphingosine kinase 1. Note that *FIZZ1* and *Ym1* gene expression is characteristic of the alternative pathway of macrophage activation.

Wang & Harris, *J Am Soc Nephrol* 22: 21–27, 2011.

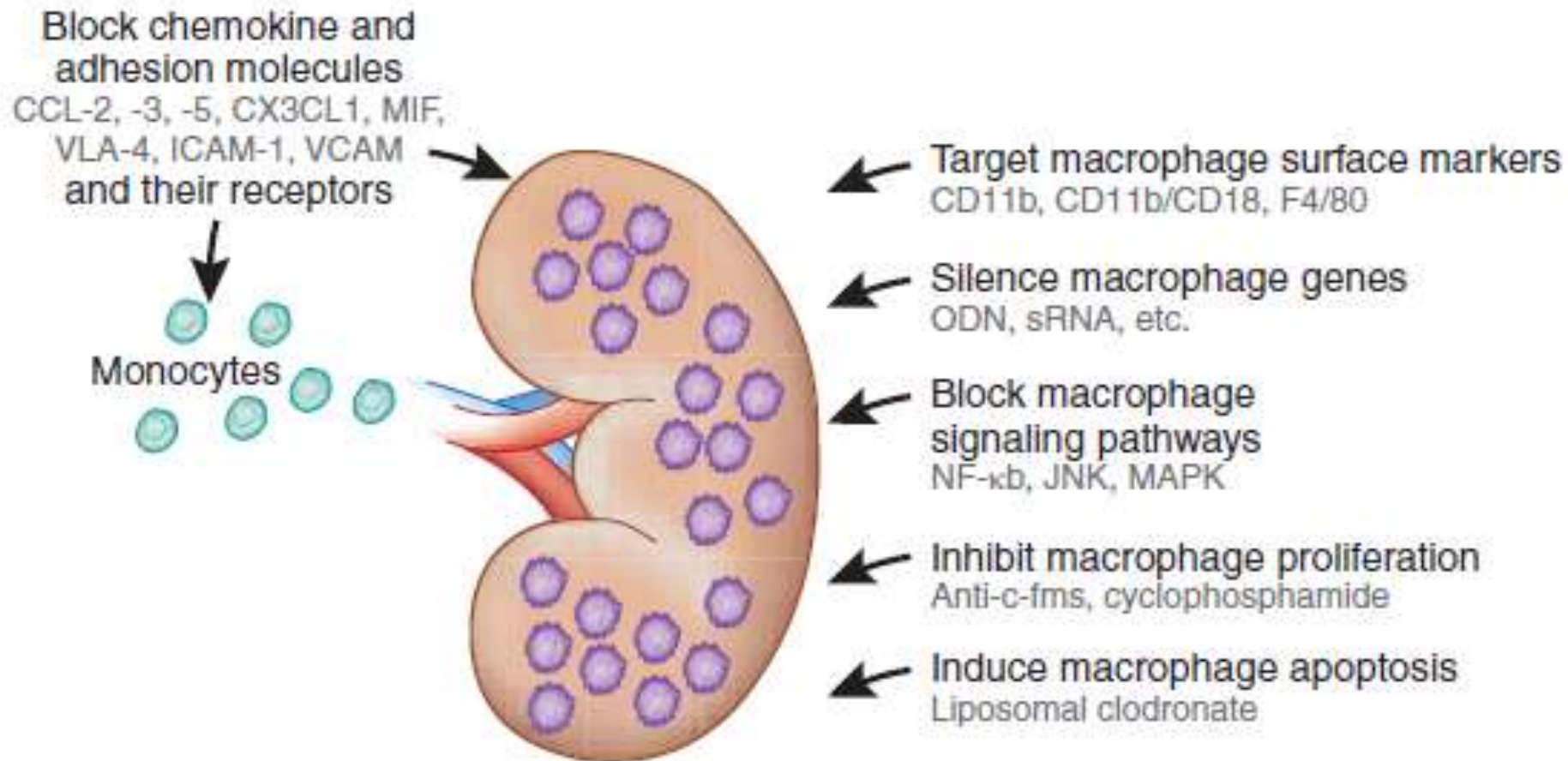
# MACROPHAGE STRATEGIES FOR TREATING RENAL DISEASE

Method	Animal Model
A. Macrophage As Target	
antimacrophage sera	Anti-GBM disease (rabbit)
ED7 antibody	AN (rat)
anti- <i>c-fms</i>	UUO (rat); diabetic nephropathy (db/db mouse)
liposomal clodronate	Anti-GBM disease
CD11b-DTR	Crescentic glomerulonephritis (mouse)
anti-CX3CR1	Crescentic glomerulonephritis (WKY rat)
anti-CCL2	Anti-GBM disease (rat and WKY rat); crescentic nephritis (mouse); anti-thy 1.1 nephritis (mouse)
CCL2 and CCL5 DNA vaccine	AN (rat)
modified CCL2 DNA vaccine	AN (rat)
anti-CCL5	Anti-thy 1.1 nephritis (mouse); nephrotoxic nephritis (mouse); immune-complex nephritis (mouse)
anti-ICAM-1, anti-CD18	Nephrotoxic nephritis (WKY rat)
NF- $\kappa$ B decoy ODN	Crescentic glomerulonephritis (rat)
ICAM-1 ODN	Renal ischaemia-reperfusion injury (rat)
CCL2 spiegelmers	Alport nephropathy (mouse)
B. Macrophage As Tool	
alternatively activated macrophages (M2a)	AN (mouse) STZ-induced DN (BALB/c and eNOS <sup>-/-</sup> mouse)
alternatively activated macrophages (M2c)	AN (mouse)
IL-1ra transfected macrophages	Anti-GBM disease (rat); UUO (rat)
IL-4 transfected macrophages	Nephrotoxic nephritis (rat)
IL-10 transfected macrophages	Nephrotoxic nephritis (rat)
macrophages transduced with I $\kappa$ B	Nephrotoxic nephritis (rat)
AN, adriamycin nephropathy; DN, diabetic nephropathy; UUO, unilateral ureteral obstruction; WKY, Wistar-Kyoto.	

Wang & Harris, *J Am Soc Nephrol* 22: 21–27, 2011.

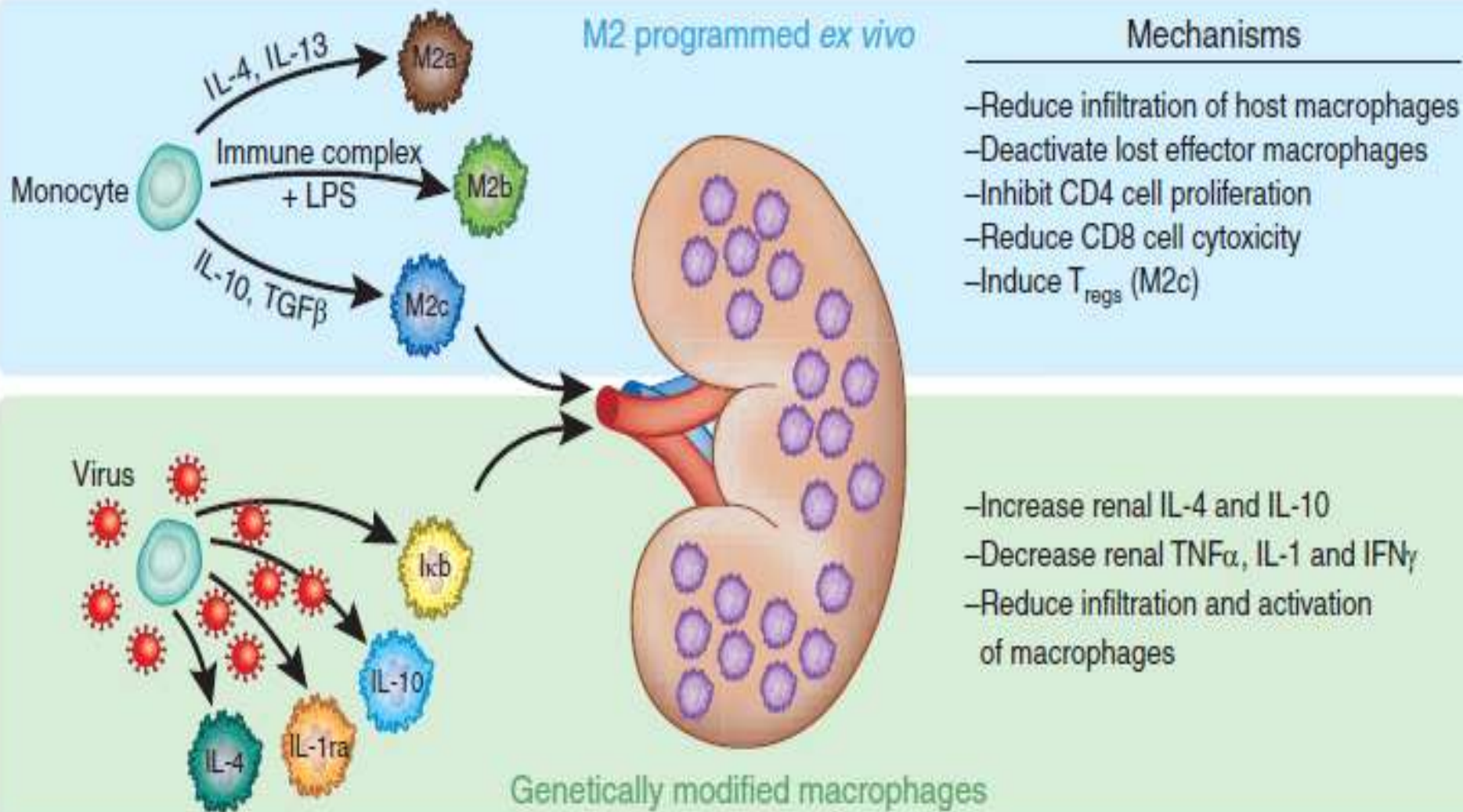


# MACROPHAGES AS A TARGET



Wang & Harris, *J Am Soc Nephrol* 22: 21–27, 2011.

# MACROPHAGES AS A TOOL



Wang & Harris, *J Am Soc Nephrol* 22: 21–27, 2011.



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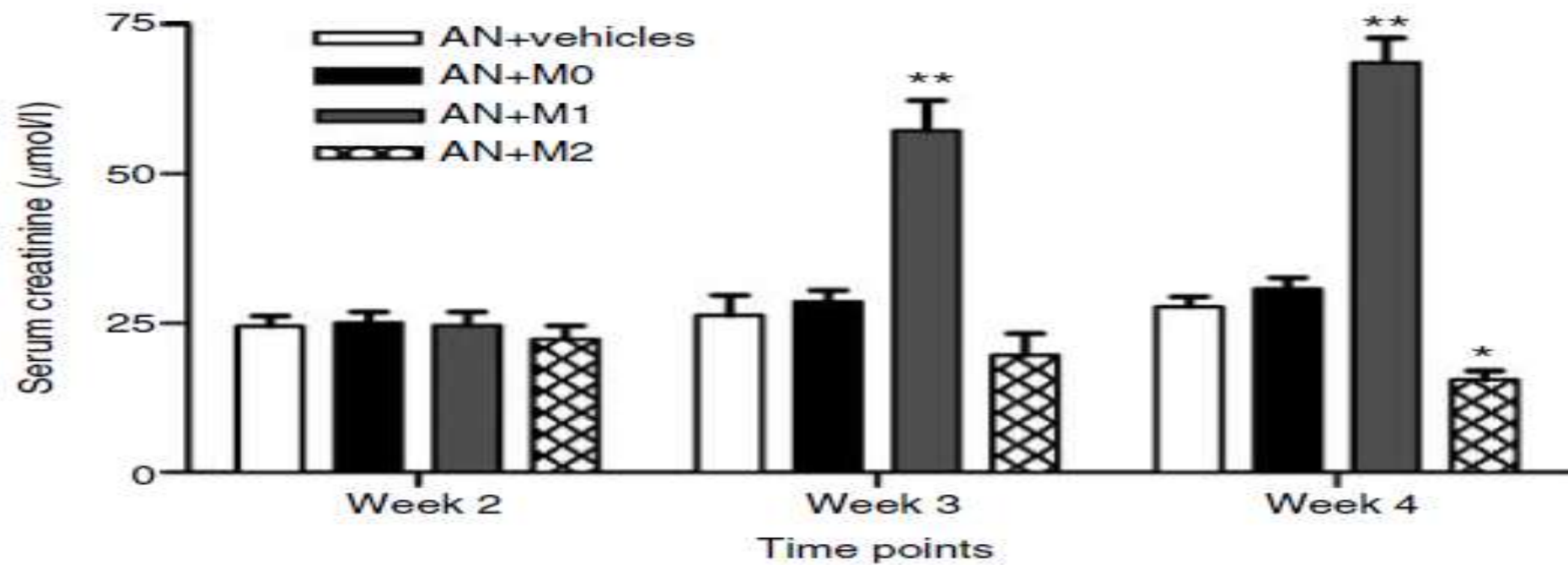
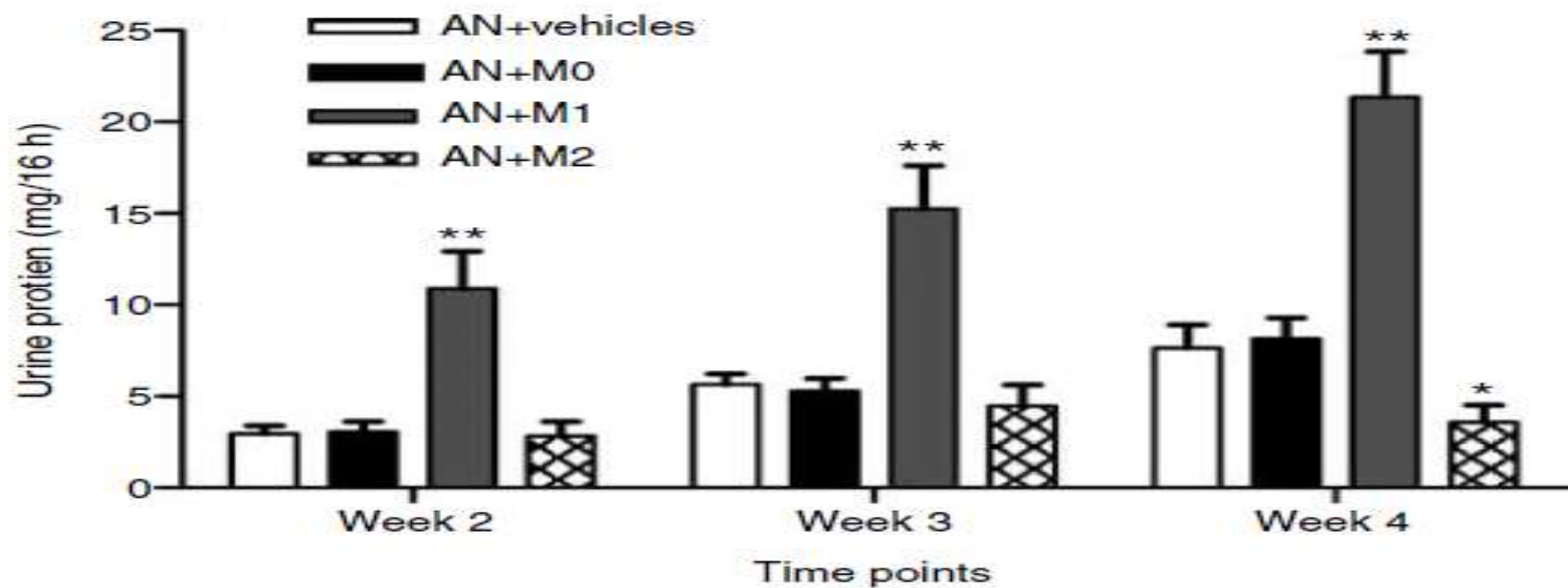
# **1. Wound-healing (IL-4/IL-13 Stimulated) (M2a) Macrophages In Adriamycin Nephropathy**

# *Ex vivo* programmed macrophages ameliorate experimental chronic inflammatory renal disease

Y Wang<sup>1</sup>, YP Wang<sup>1</sup>, G Zheng<sup>1</sup>, VWS Lee<sup>1</sup>, L Ouyang<sup>1</sup>, DHH Chang<sup>1</sup>, D Mahajan<sup>1</sup>, J Coombs<sup>1</sup>, YM Wang<sup>2</sup>, SI Alexander<sup>2</sup> and DCH Harris<sup>1</sup>

<sup>1</sup>Centre for Transplantation and Renal Research, The University of Sydney at Westmead Millennium Institute, Westmead, New South Wales, Australia and <sup>2</sup>Centre for Kidney Research, The Children's Hospital at Westmead, Westmead, New South Wales, Australia

Macrophages were isolated from the spleens of BALB/c mice and stimulated with lipopolysaccharide to induce M1 macrophages or with interleukin-4 (IL-4) and IL-13 to induce M2a macrophages. These macrophages were then infused into SCID mice with adriamycin nephropathy and examined after 2,3,4 weeks.





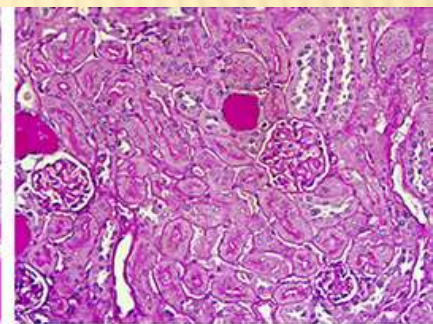
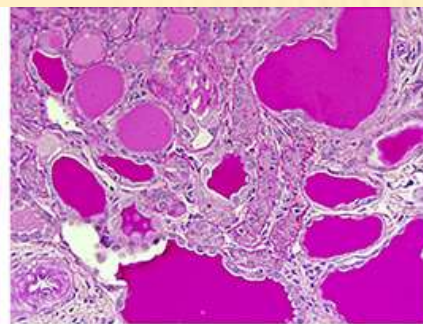
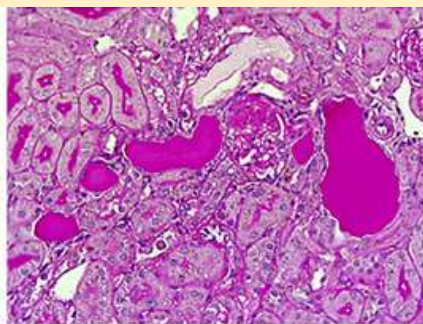
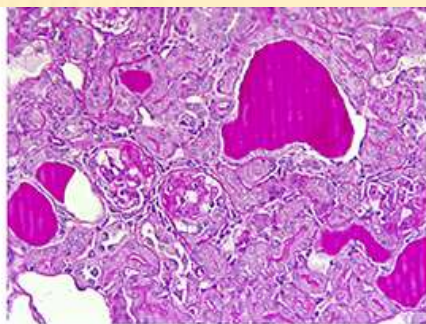
AN + Vehicle

AN + M0

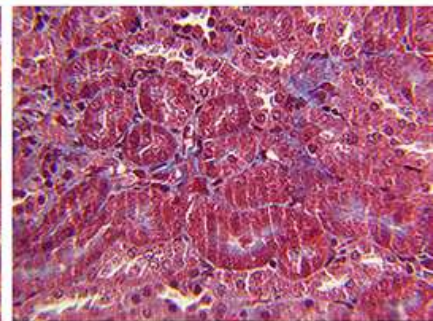
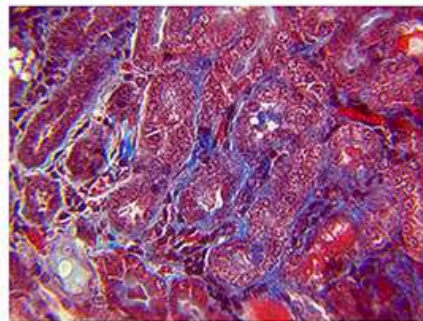
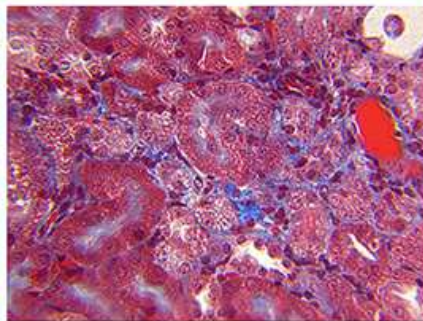
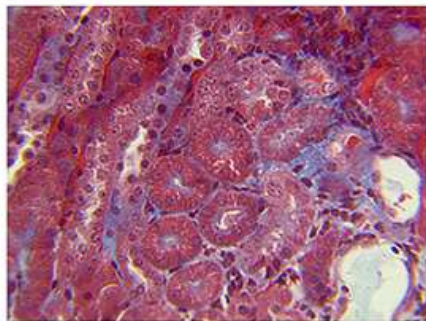
AN + M1

AN + M2

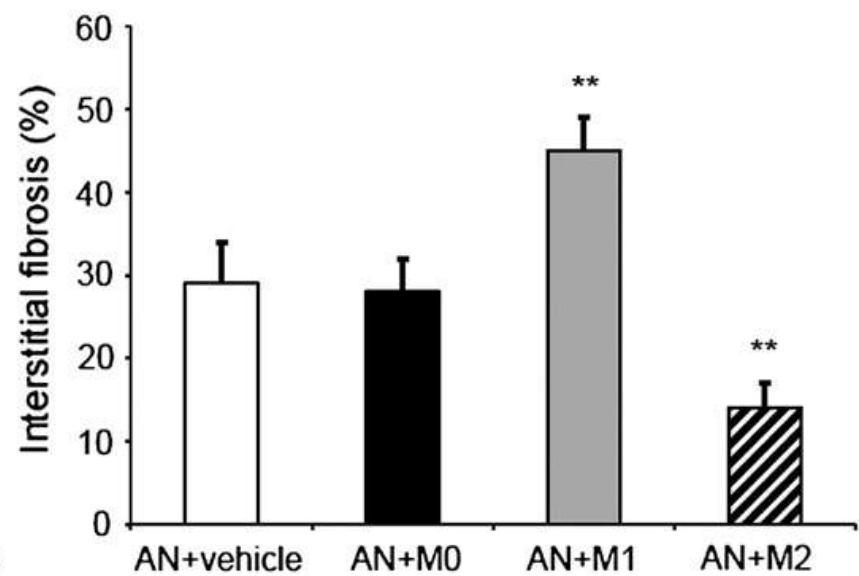
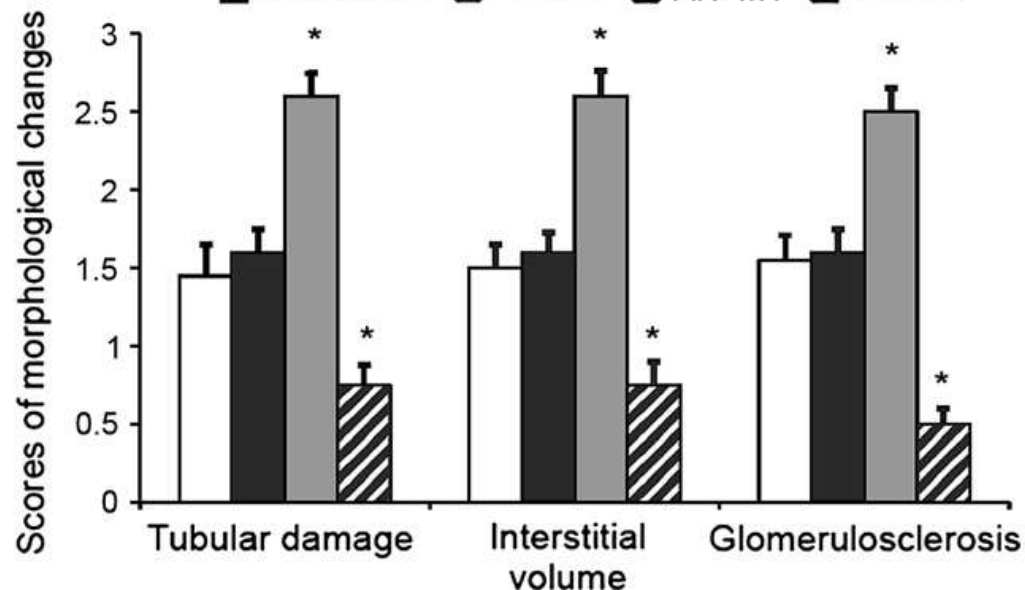
PAS



Trichrome



■ AN+vehicle ■ AN+M0 ■ AN+M1 ■ AN+M2



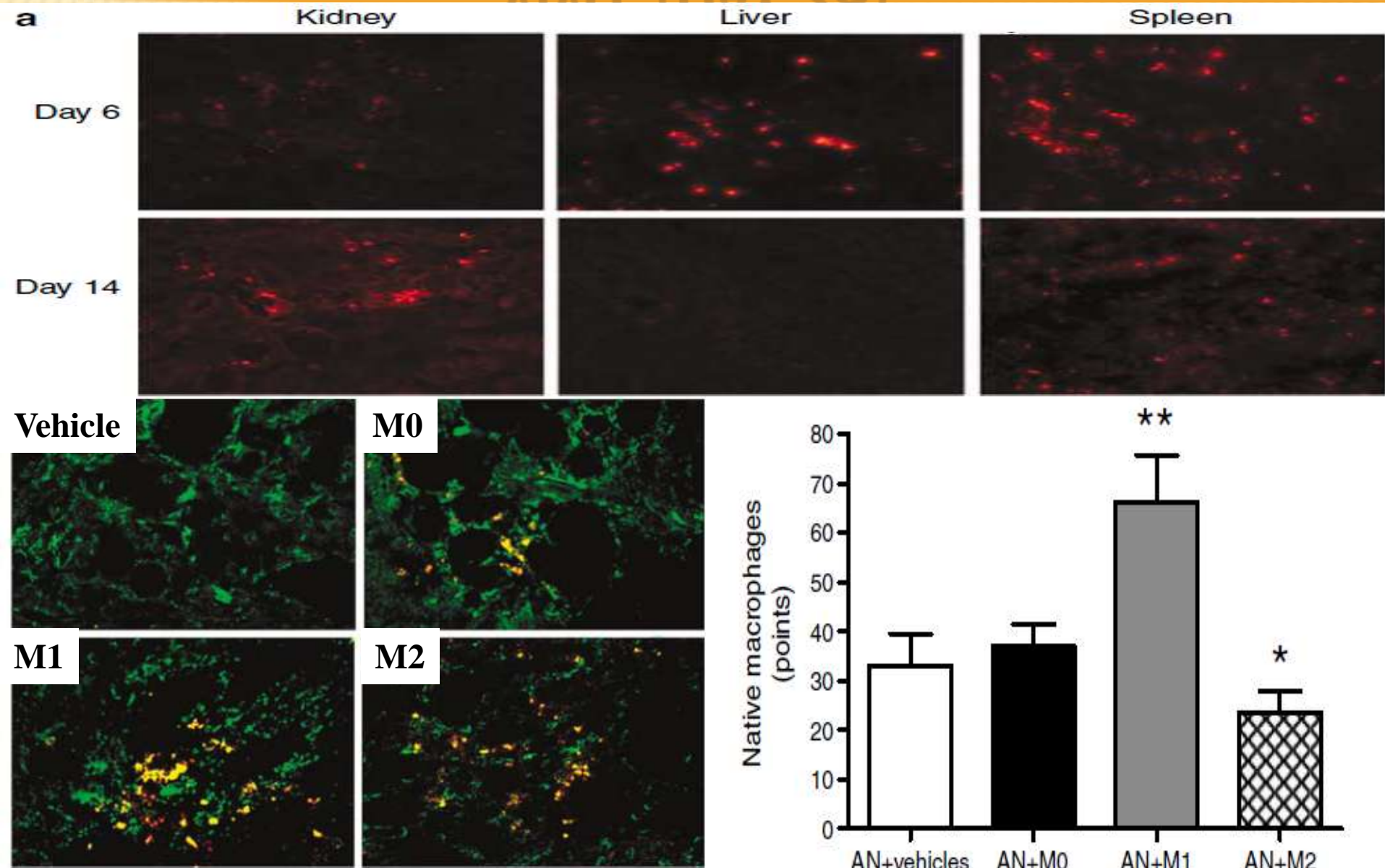


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# TRACKING OF TRANSFUSED MACROPHAGES

## ????????????

# TRACKING OF TRANSFUSED MACROPHAGES IN VIVO (DAY 28)



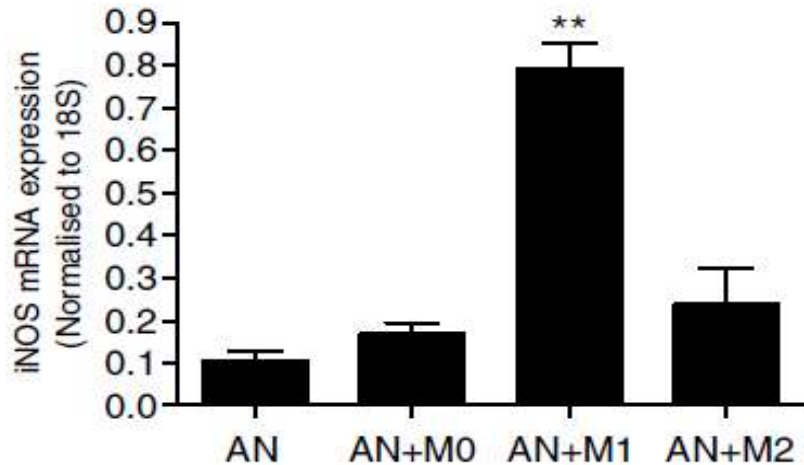
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# STABILITY OF TRANSFUSED MACROPHAGES

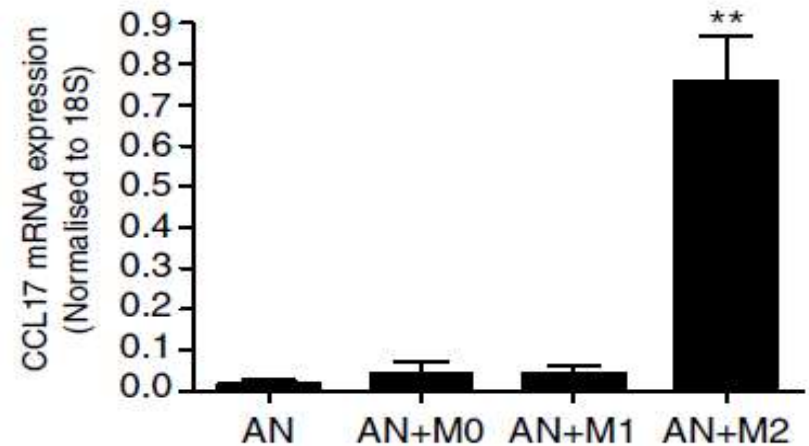
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# STABILITY OF TRANSFUSED MACROPHAGES IN VIVO AT DAY 28

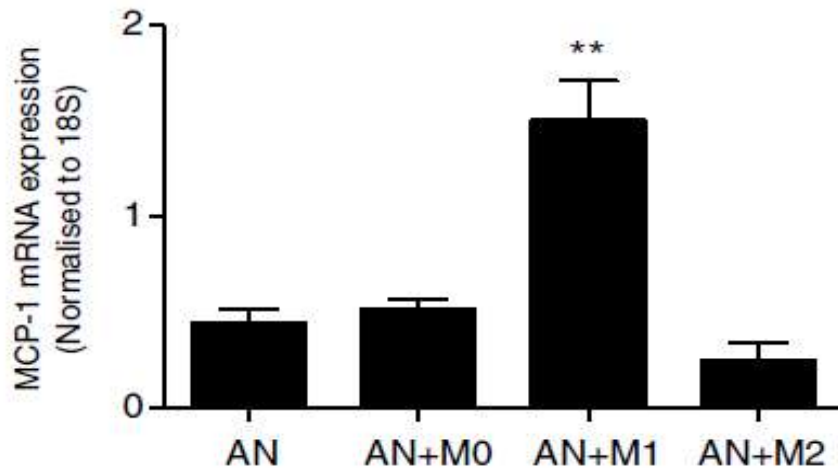
Macrophage iNOS



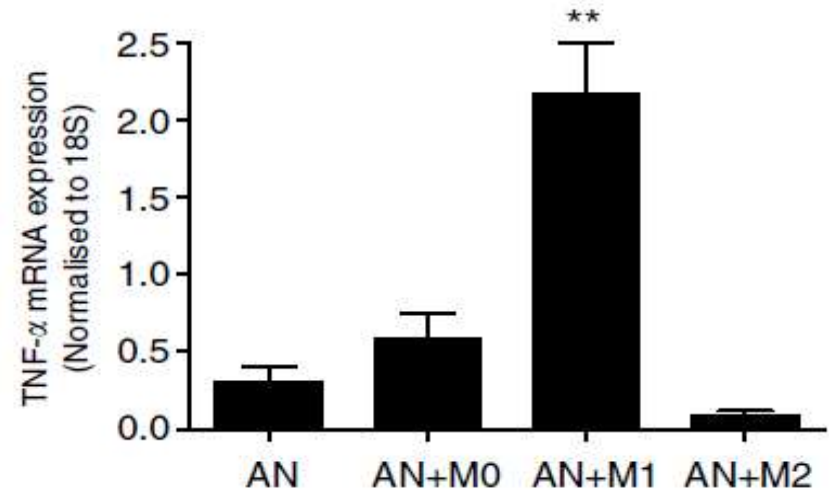
Macrophage CCL17



Macrophage CCL2



Macrophage TNF- $\alpha$





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## **2. Regulatory (IL-10/TGF- $\beta$ stimulated) (M2c) Macrophages In Adriamycin Nephropathy**

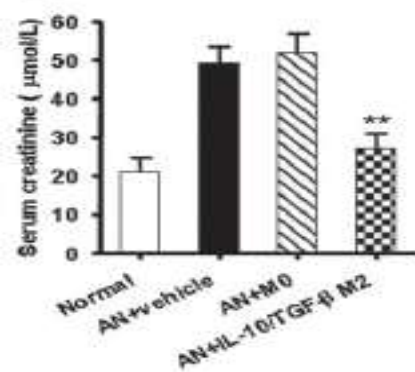
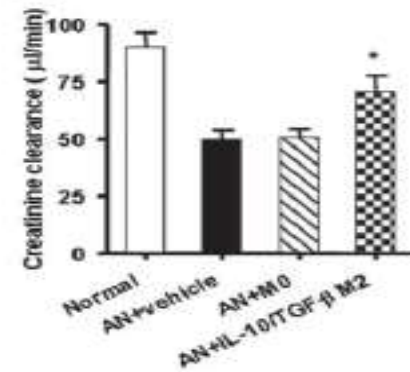
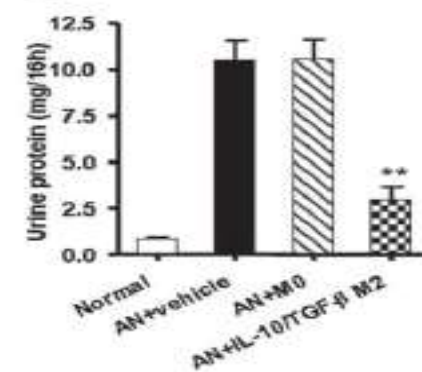
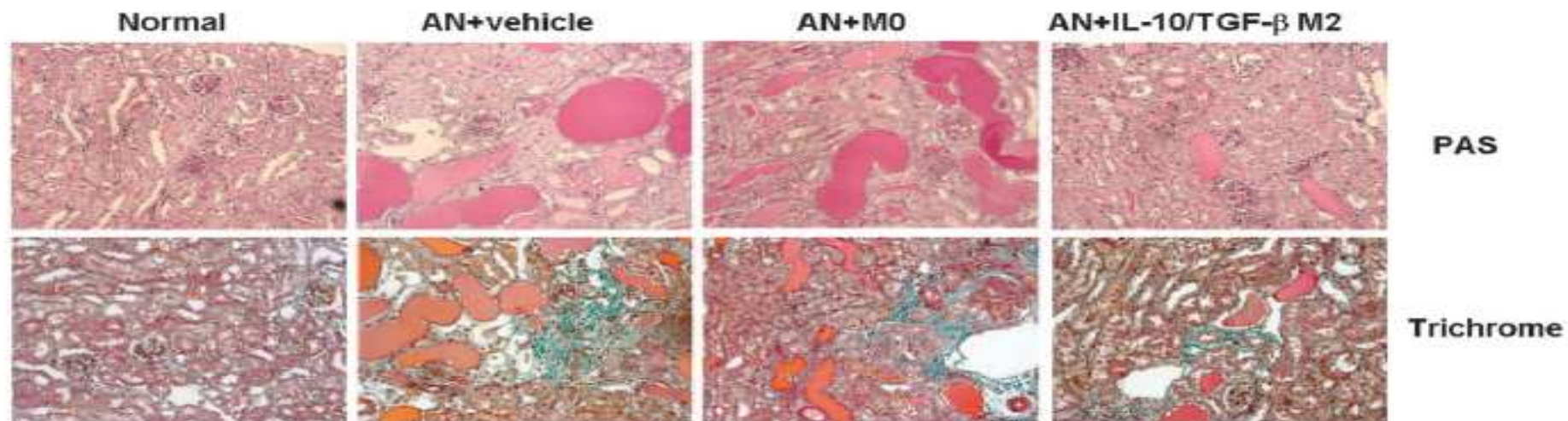
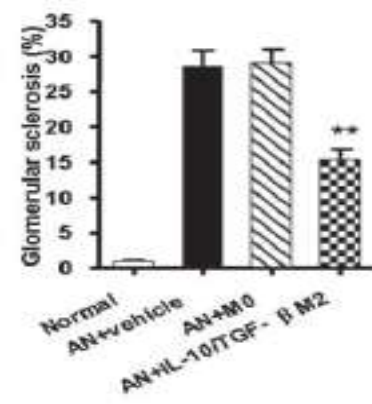
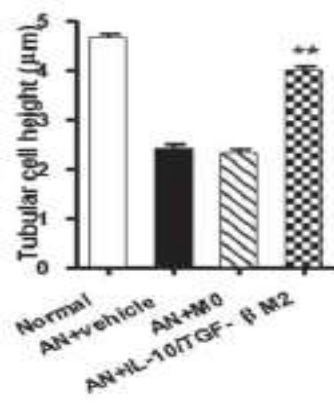
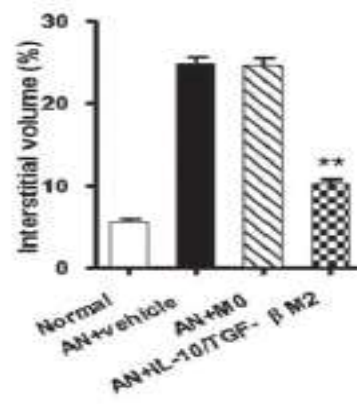
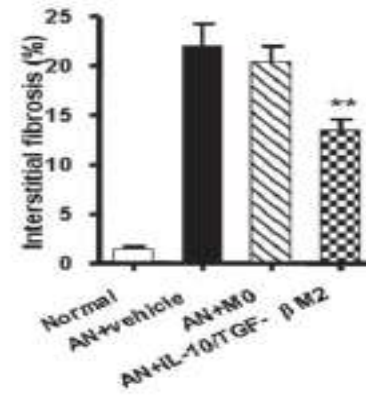
# IL-10/TGF- $\beta$ -Modified Macrophages Induce Regulatory T Cells and Protect against Adriamycin Nephrosis

Qi Cao,\* Yiping Wang,\* Dong Zheng,\* Yan Sun,\* Ya Wang,\* Vincent W.S. Lee,\*  
Guoping Zheng,\* Thian Kui Tan,\* Jon Ince,\* Stephen I. Alexander,<sup>†</sup> and David C.H. Harris\*

\*Centre for Transplantation and Renal Research, University of Sydney, Westmead Millennium Institute, Sydney, New South Wales, Australia; and <sup>†</sup>Centre for Kidney Research, Children's Hospital at Westmead, Sydney, New South Wales, Australia

*J Am Soc Nephrol* 21: 933–942, 2010. doi: 10.1681/ASN.2009060592

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**A****B****C****D****E****F****H****I**



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# **REGULATORY EFFECT OF M2c MACROPHAGES**

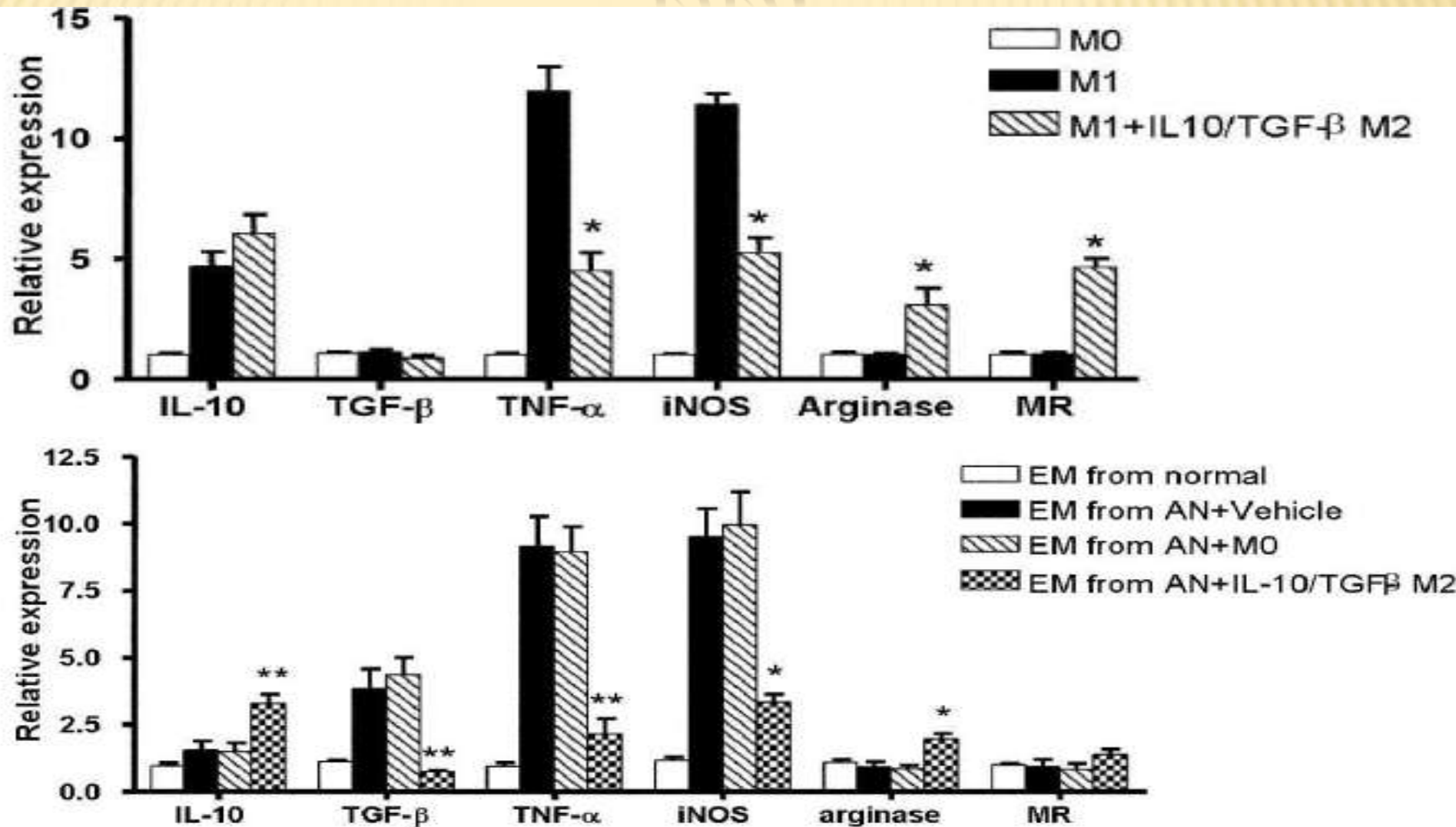
# EFFECT OF M2c MACROPHAGES ON ENDOGENOUS INFLAMMATORY CELLS

Parameter	AN+Vehicle	AN+M0	AN+IL-10/ TGF- $\beta$ M2
CD4 <sup>+</sup> T cells	72.3 $\pm$ 13.6	70.3 $\pm$ 15.7	25.3 $\pm$ 7.2 <sup>a</sup>
CD8 <sup>+</sup> T cells	36.3 $\pm$ 5.5	38.1 $\pm$ 4.7	17.6 $\pm$ 5.6 <sup>a</sup>
Macrophages	64.3 $\pm$ 10.8	66.4 $\pm$ 10.1	42.9 $\pm$ 8.4 <sup>a</sup>

Data are means  $\pm$  SEM of cells per  $\times 400$  field.

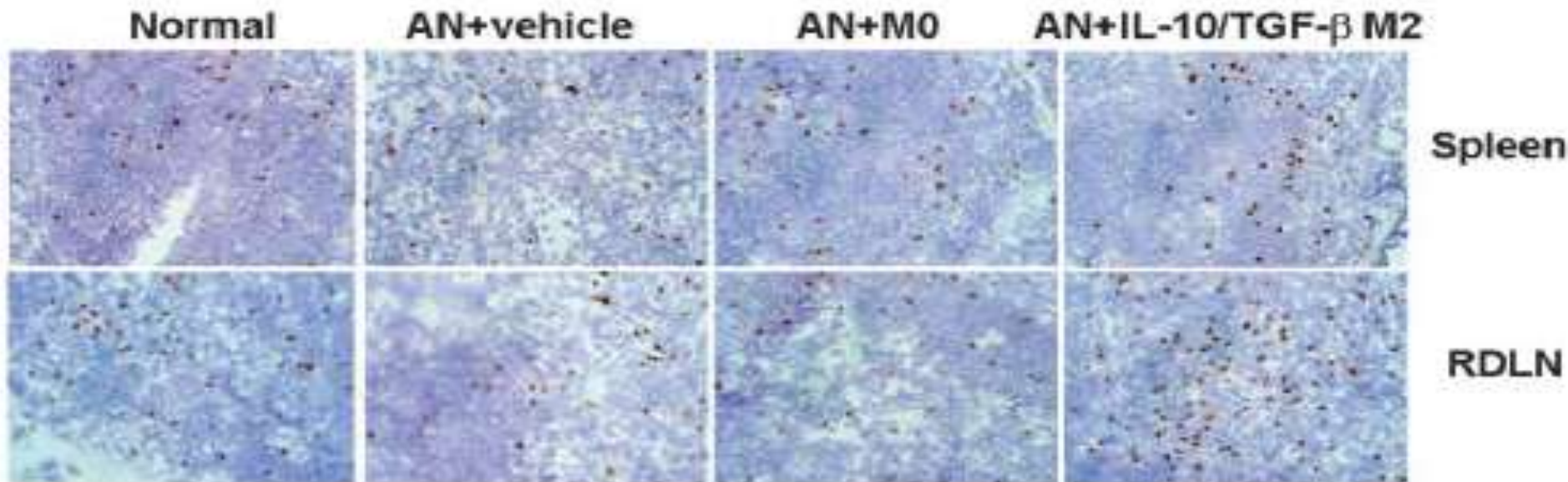
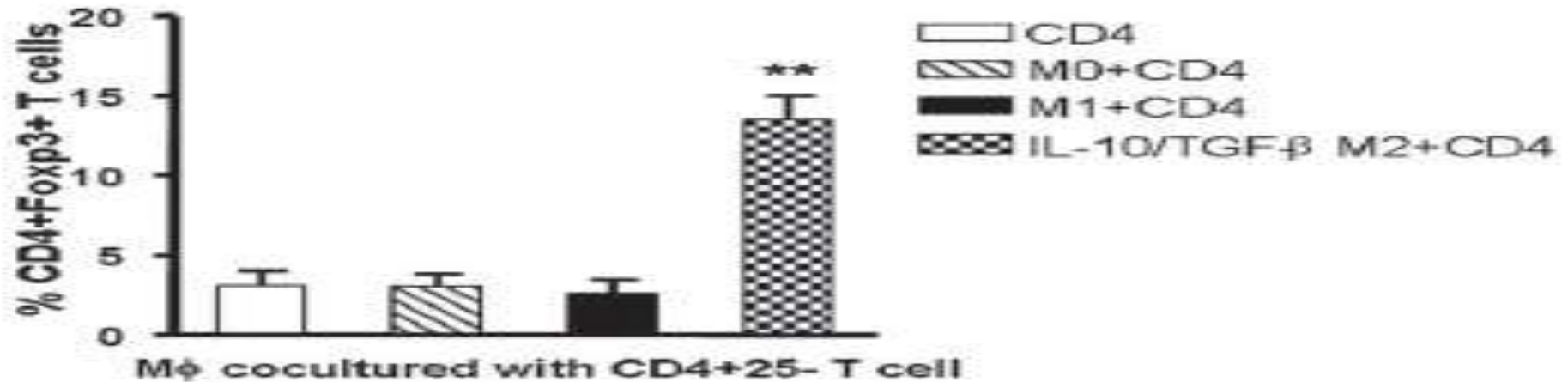
<sup>a</sup> $P < 0.01$  versus AN+M0.

# M2c SUPPRESSES M1 *IN VITRO* AND *ENDOGENOUS RENAL* MACROPHAGES *IN VIVO*





# M2c MACROPHAGES INDUCES Tregs IN VITRO & IN VIVO

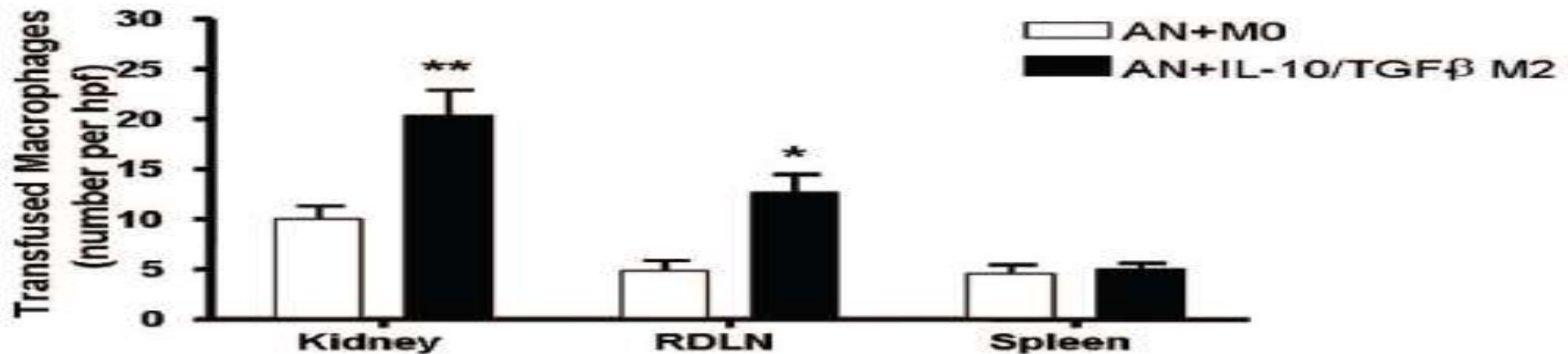
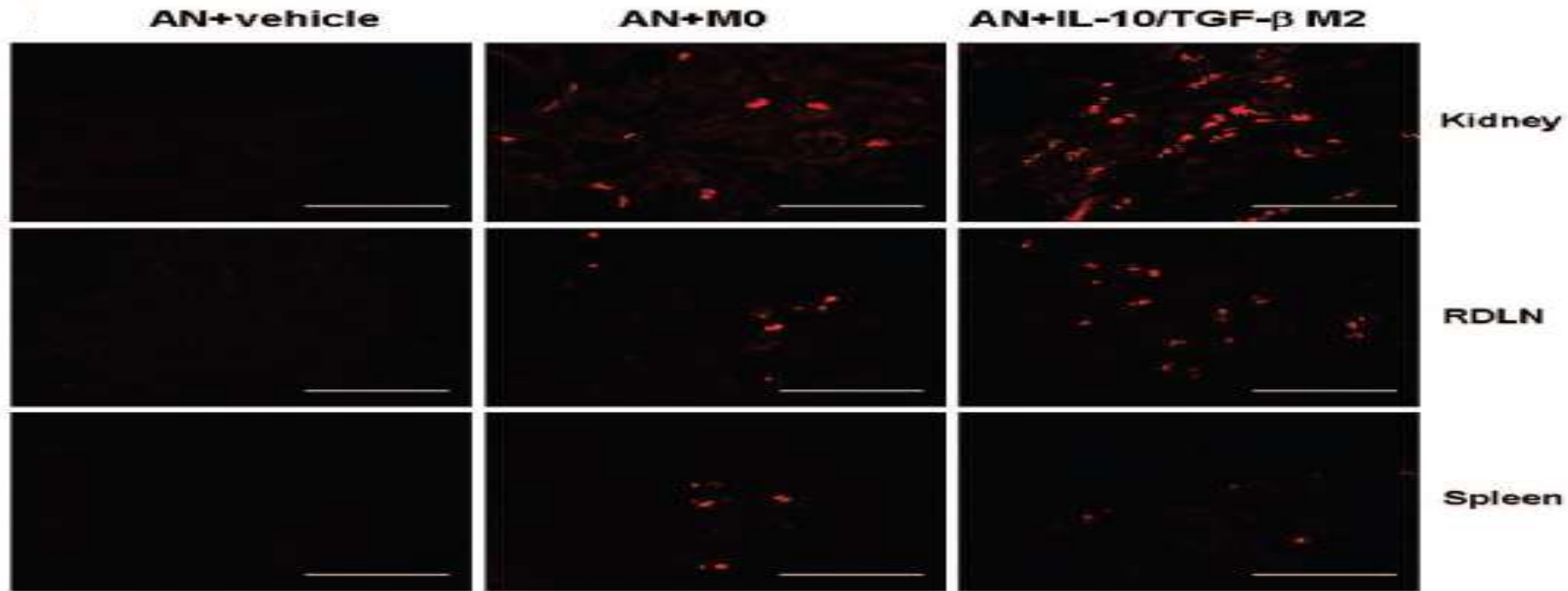


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# TRACKING OF TRANSFUSED MACROPHAGES

## ????????????

# SELECTIVE TRACKING OF TRANSFUSED M2c (28 days)



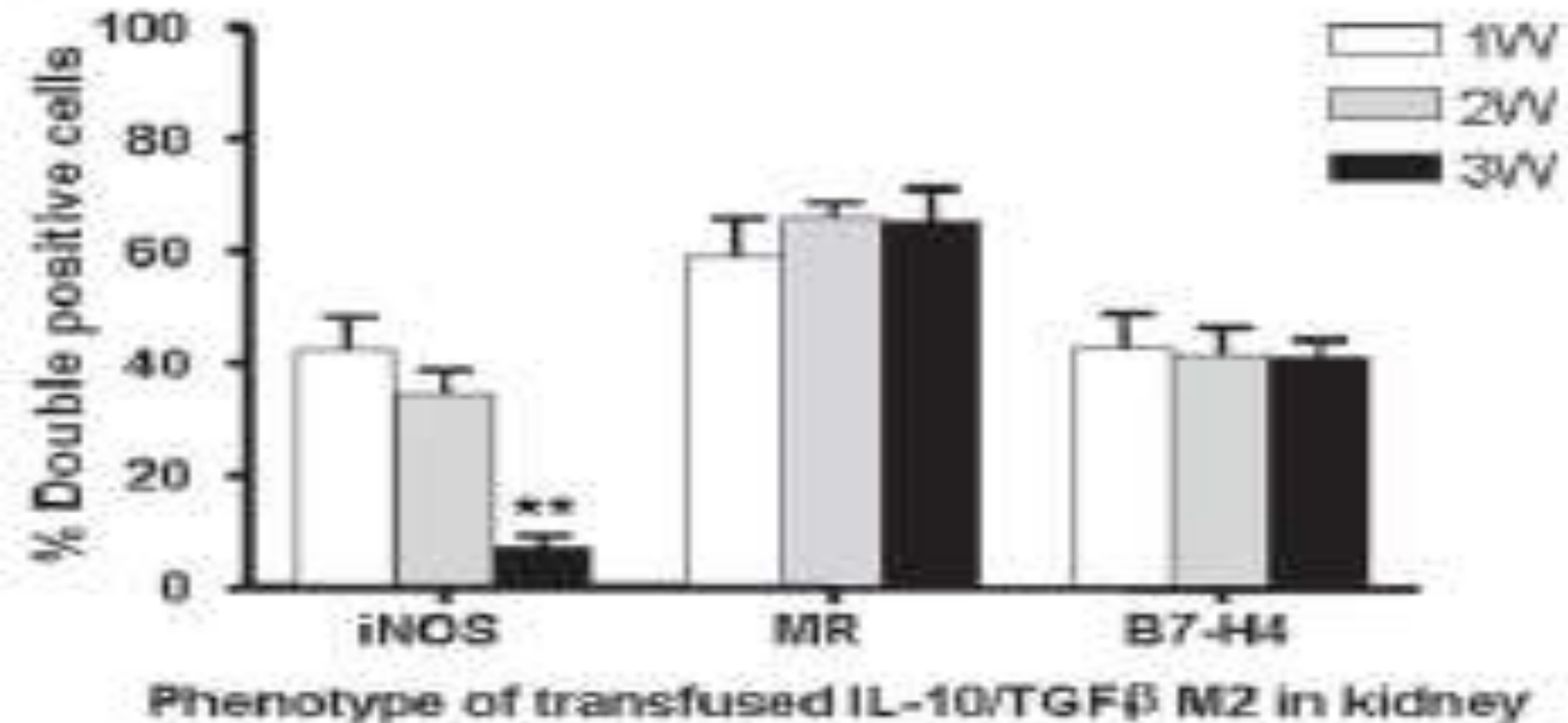


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# **STABILITY OF TRANSFUSED MACROPHAGES**

**??????????**

# MAINTENANCE OF ANTI-INFLAMMATORY PHENOTYPES *IN VIVO* OF TRANSFUSED M2c



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### **3. Wound-healing (M2a) Macrophages in Diabetic Nephropathy**



CTRR

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and Renal Research



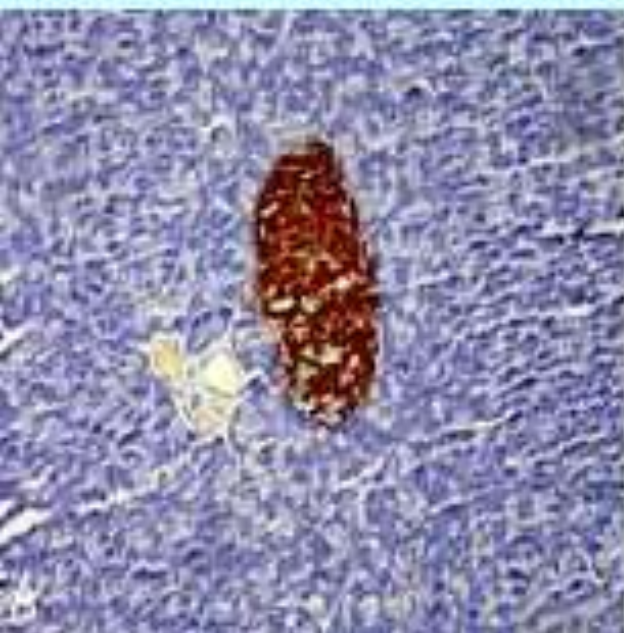
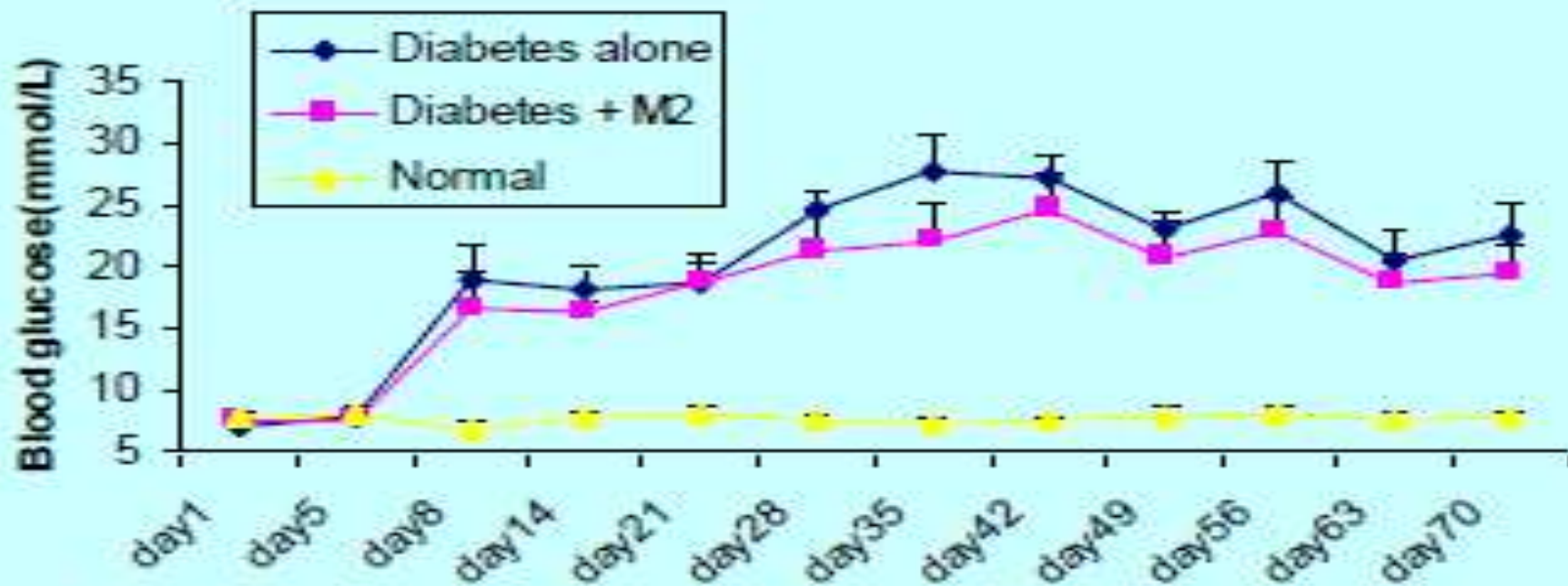
# Regulatory macrophages protect against renal injury in murine streptozotocin-induced diabetes



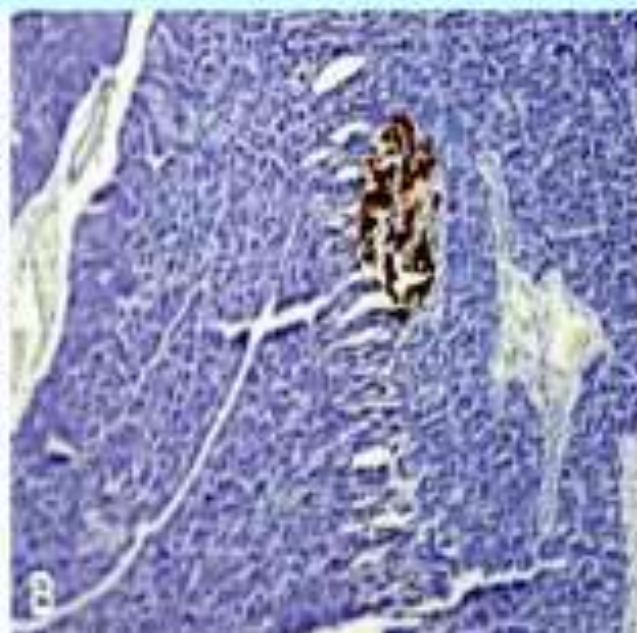
D Zheng, YP Wang, VWS Lee, Q Cao, G Zheng, Y Sun, Y Wang, SI Alexander<sup>1</sup> and DCH Harris

Centre for Transplant and Renal Research, The University of Sydney, Westmead Millennium Institute, Sydney, NSW, Australia and <sup>1</sup>Centre for Kidney Research, Children's Hospital at Westmead, Sydney, NSW, Australia.

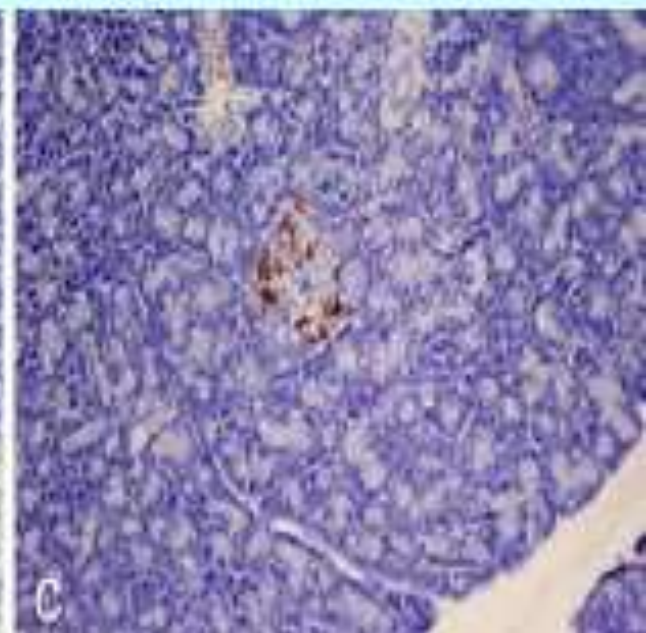
**Macrophages were separated from splenocytes of BALB/c mice and cultured with IL-4 and IL-13. Mice underwent adoptive transfer with  $1 \times 10^6$  M2/per mouse, followed by tail injection with two doses of STZ (75mg/kg and 150mg/kg). Blood glucose levels were monitored daily. Mice were sacrificed at the 10<sup>th</sup> week after STZ injection. Renal function and histopathological injury were assessed.**



**Normal**



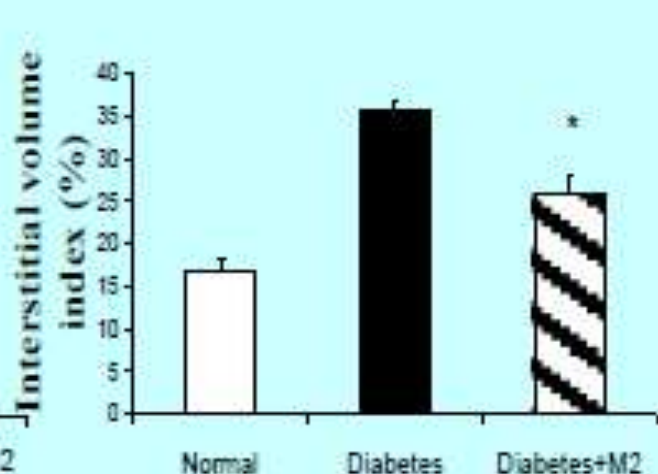
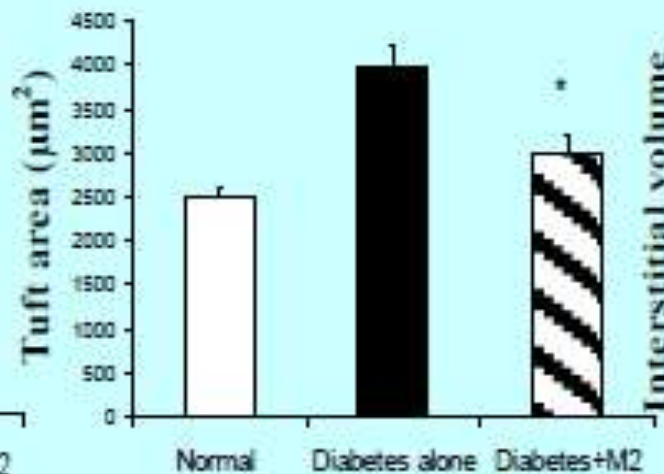
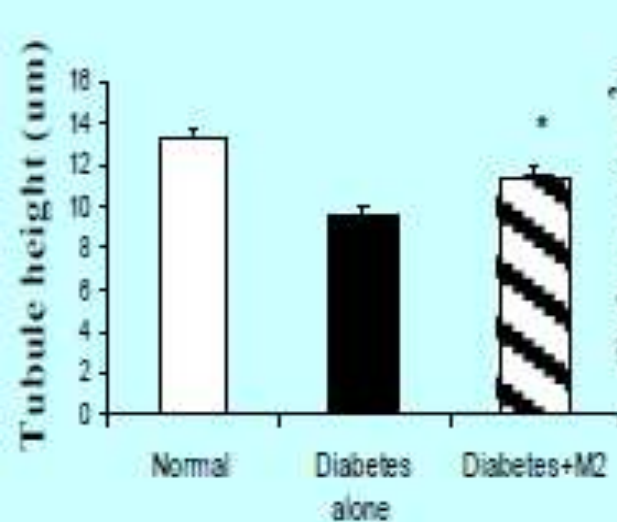
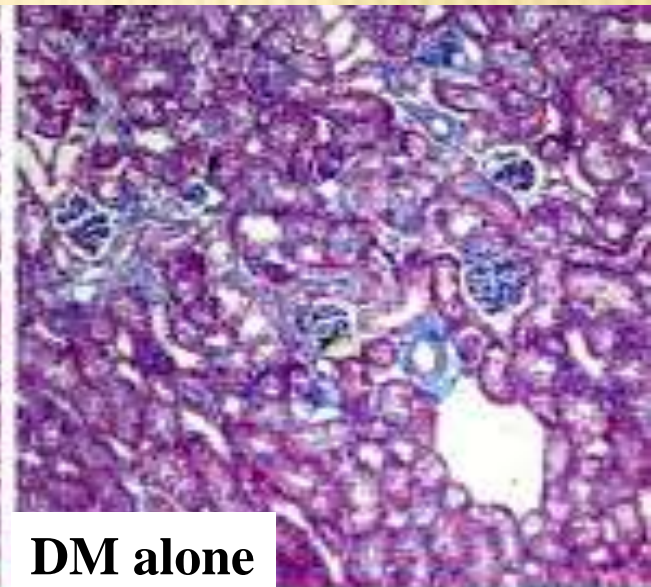
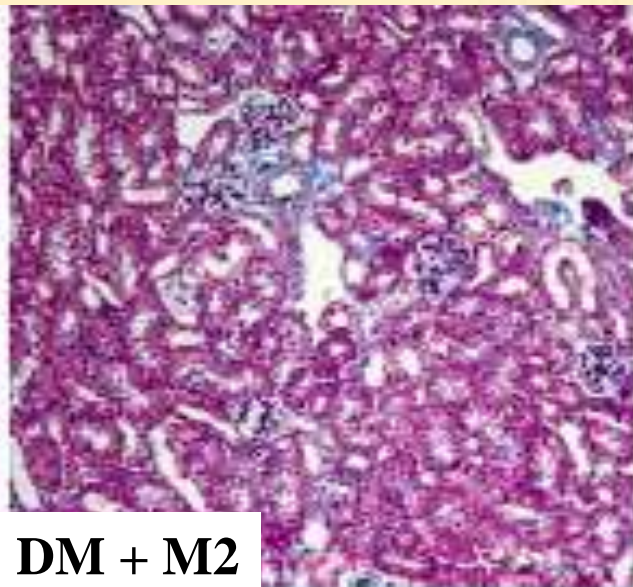
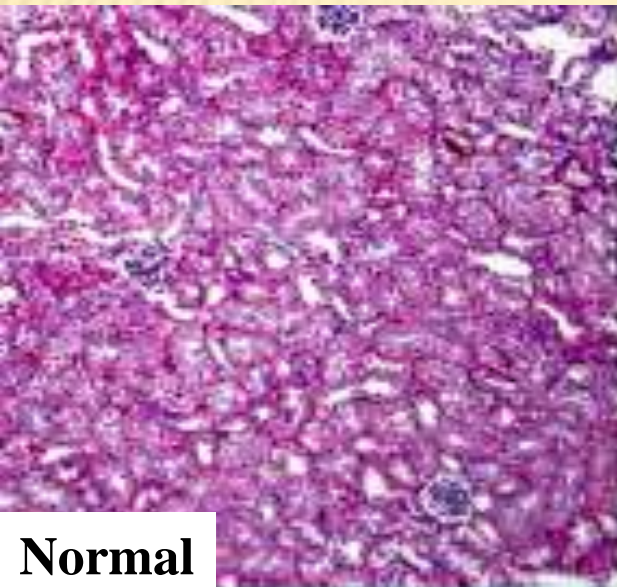
**DM + M2**



**DM alone**

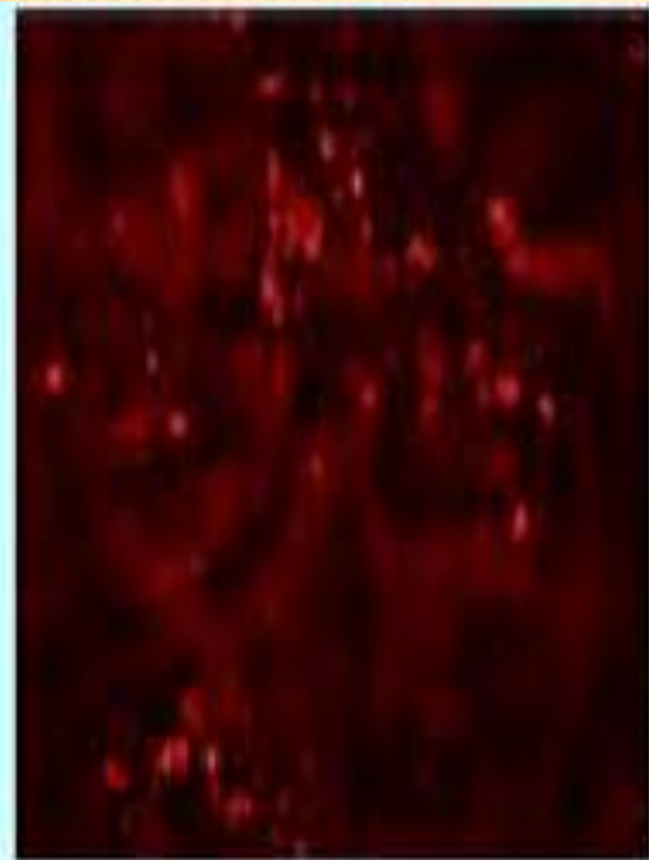


# M2a MACROPHAGES AMELIORATES RENAL FIBROSIS.





# ***IN VIVO TRACKING OF TRANSFERRED MACROPHAGES 10 WEEKS***



**Kidney**



**Pancreas**



**Spleen**

---

# **CAN M2a AMELIORATE RENAL INJURY IN ESTABLISHED DIABETIC NEPHROPATHY**

**????????????**

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**M2a**

**VS**

**M2c**



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# **BMD VS SPLENIC MACROPHAGES**

# OBSTACLES FOR USE IN HUMAN

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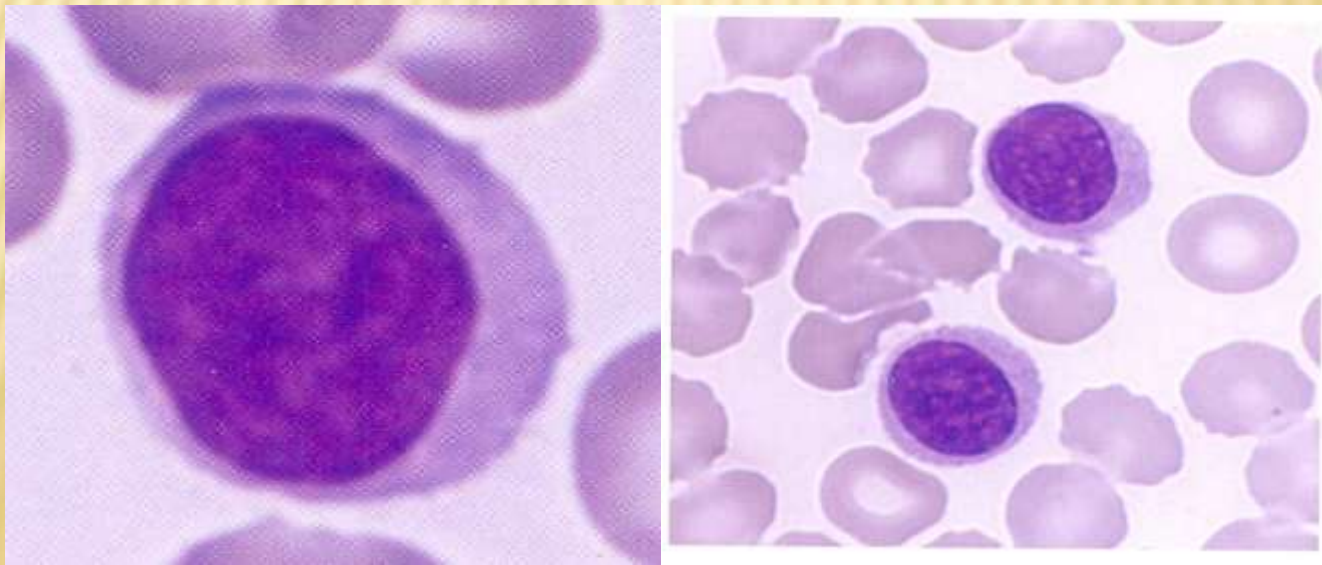
- Adequate purity and sufficient numbers of macrophages from humans for adoptive transfer.
- Dose, timing and frequency of use of regulatory cells.
- Immunosuppression: malignancy & infection.
- Stability and possibility of phenotypic switch.
- Lack of human studies.
- The effect of M2 macrophages in advanced disease is unproven.

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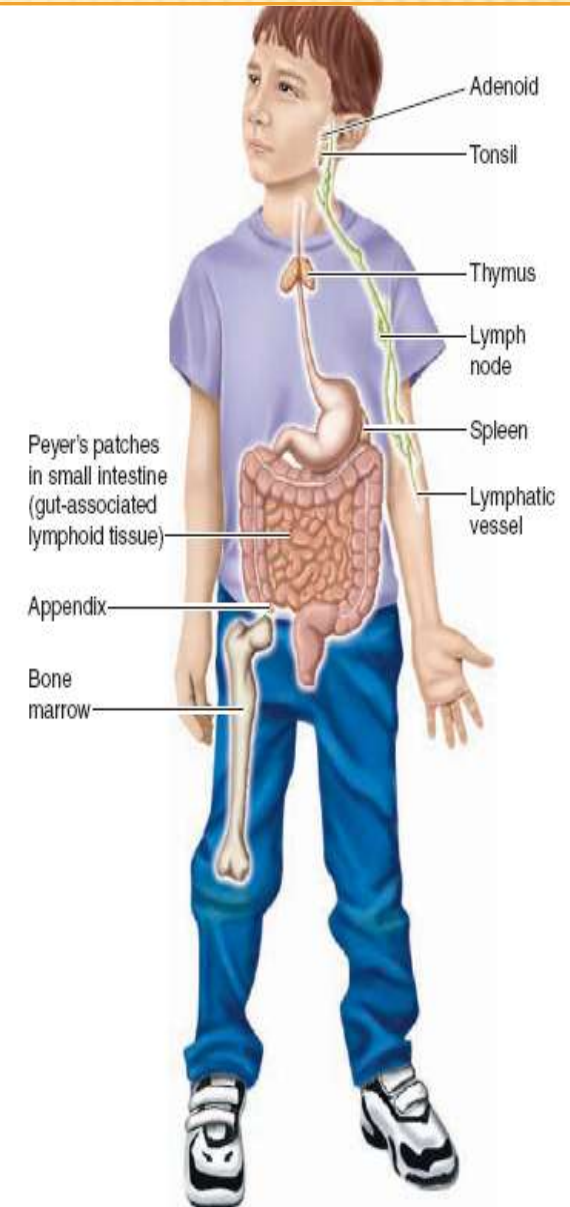
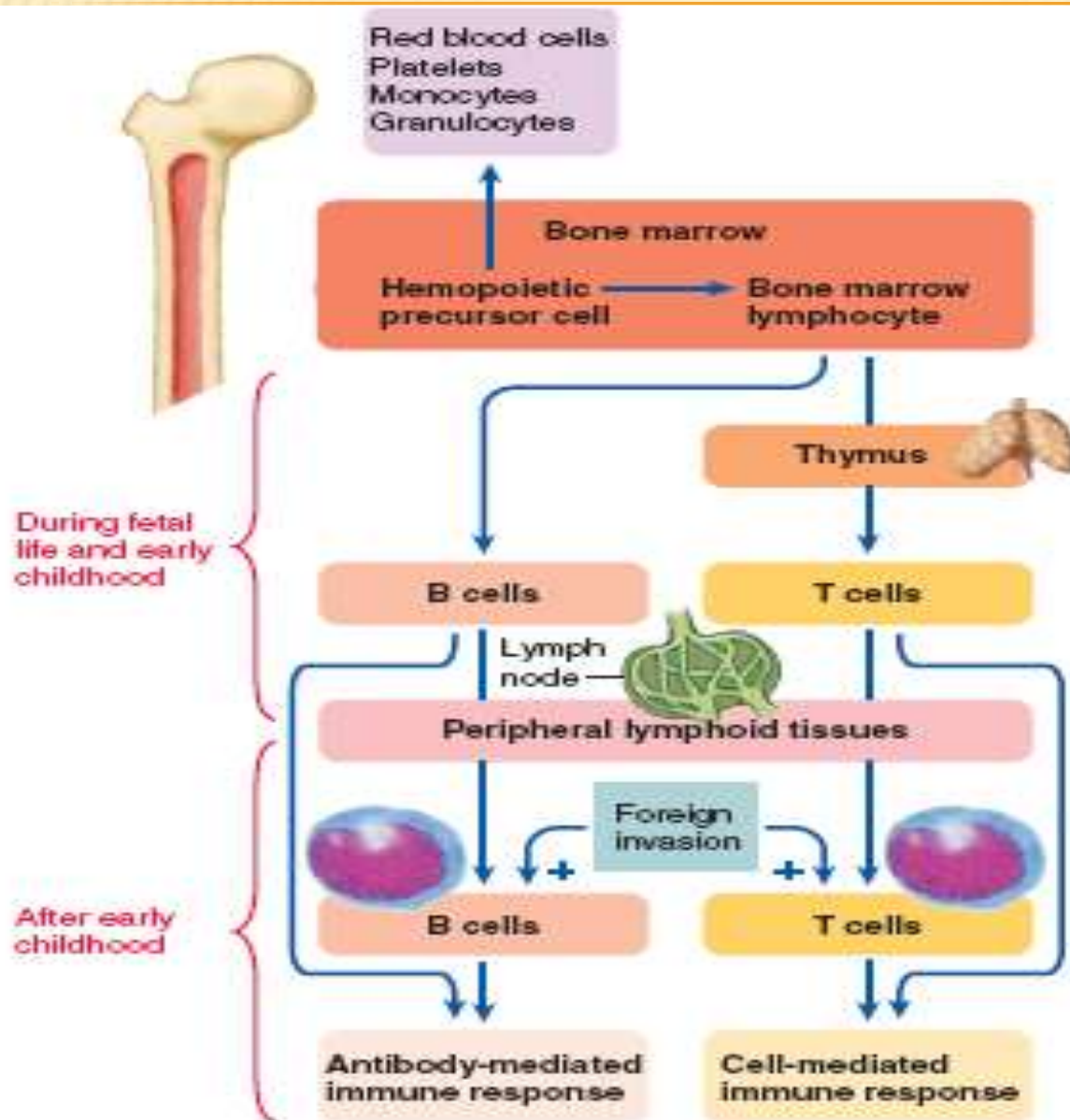
# LYMPHOCYTES

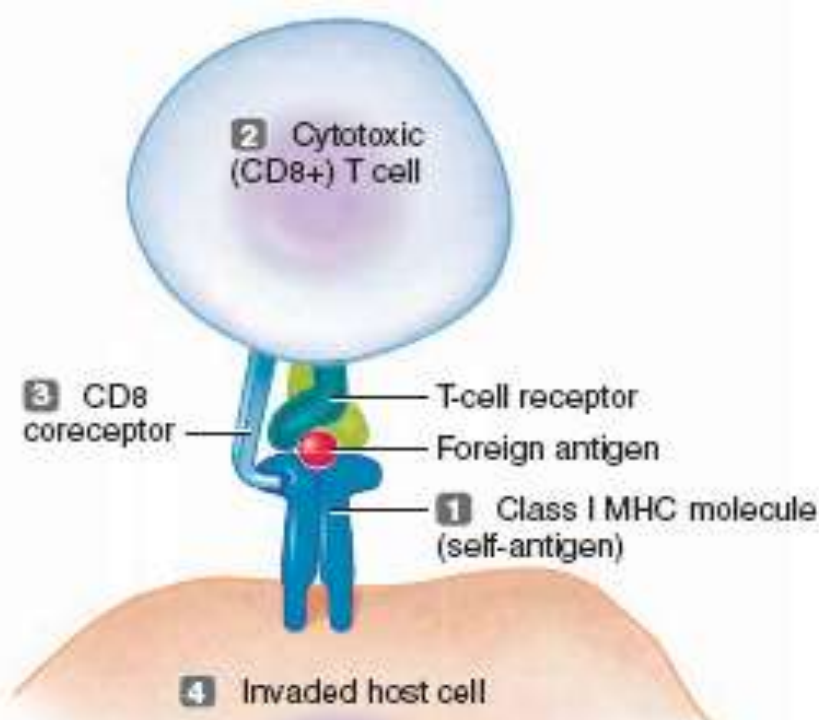


- ✗ T lymphocytes: cause direct destruction of virus invaded cells and mutant cells (Cell mediated immunity)
- ✗ B lymphocytes: secrete antibodies that indirectly lead to the destruction of foreign material (Humoral immunity)



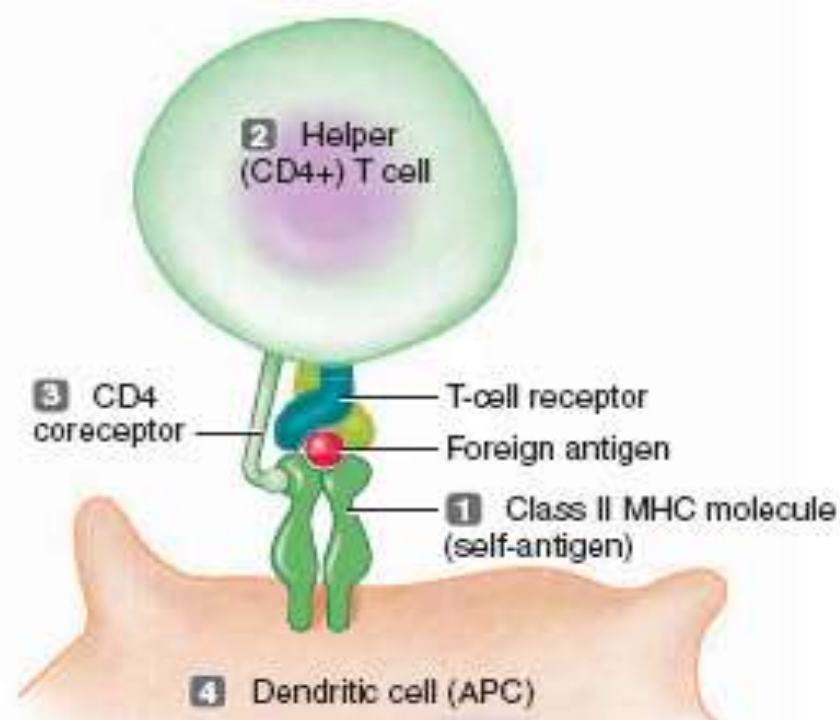
# ORIGIN OF T AND B LYMPHOCYTES





- 1** Class I MHC molecules are found on surface of all cells.
- 2** They are recognized only by cytotoxic (CD8+) T cells.
- 3** CD8 coreceptor links the two cells together.
- 4** Linked in this way, cytotoxic T cells can destroy body cells if invaded by foreign (viral) antigen.

**(a) Class I MHC self-antigens**

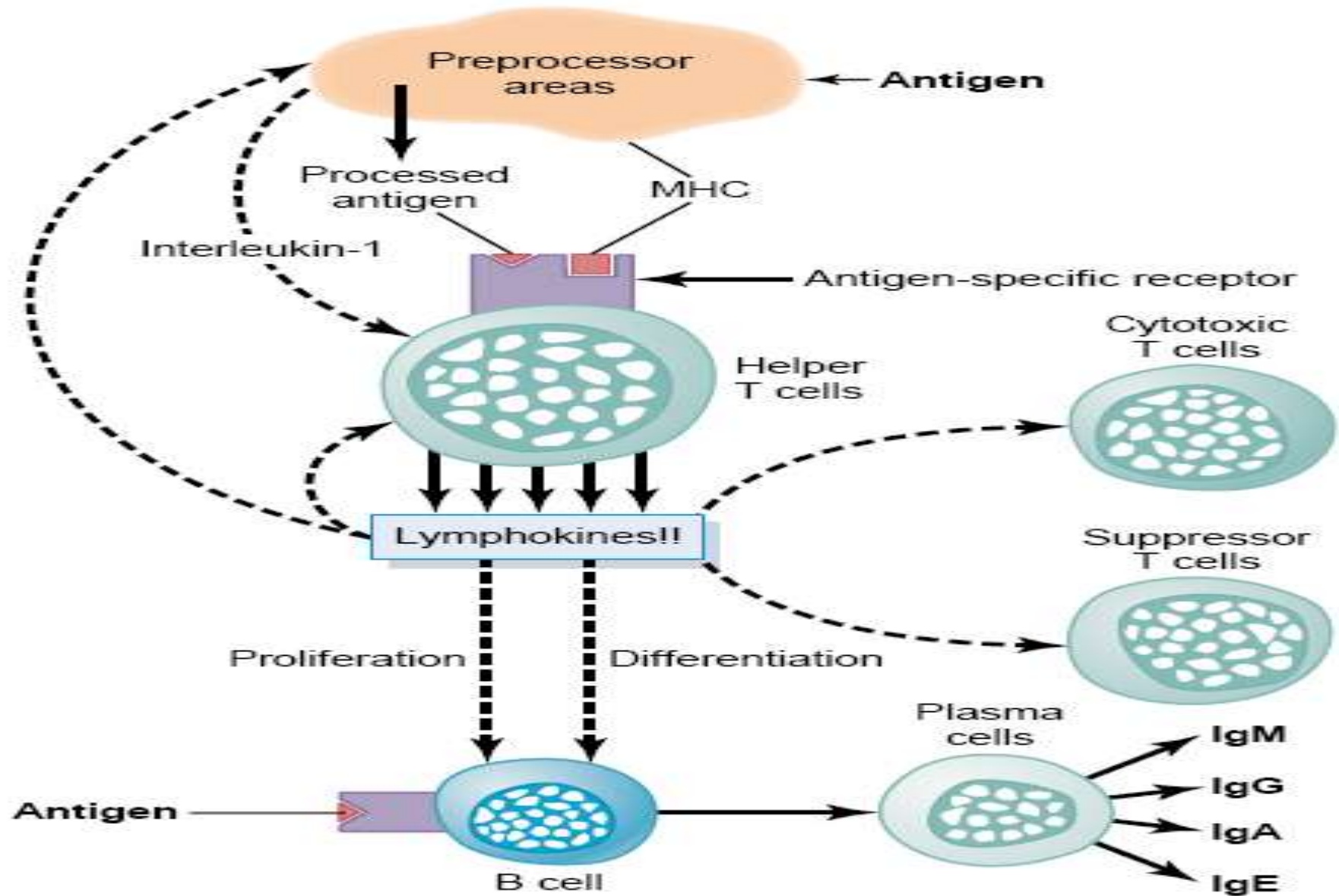


- 1** Class II MHC molecules are found on the surface of immune cells with which helper T cells interact: dendritic cells, macrophages, and B cells.
- 2** They are recognized only by helper (CD4+) T cells.
- 3** CD4 coreceptor links the two cells together.
- 4** To be activated, helper T cells must bind with a class II MHC-bearing APC (dendritic cell or macrophage). To activate B cells, helper T cell must bind with a class II MHC-bearing B cell with displayed foreign antigen.

**(b) Class II MHC self-antigens**



# PIVOTAL ROLE OF HELPER T CELL



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# **ROLE OF LYMPHOCYTES IN ISCHEMIC AKI (IRI)**

- 
- ✗ T cells were not expected to participate in the initial renal injury based on traditional ideas about the immunological functions of T cells.



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# **LYMPHOCYTES ROLE IN EARLY & LATE RENAL INJURY**

# The Role of the B7 Costimulatory Pathway in Experimental Cold Ischemia/Reperfusion Injury

Moriatsu Takada,\* Anil Chandraker,<sup>†</sup> Kari C. Nadeau,<sup>§</sup> Mohamed H. Sayegh,<sup>†</sup> and Nicholas L. Tilney\*

*\*Surgical Research Laboratory and Department of Surgery, and <sup>†</sup>Renal Division, Department of Medicine, Brigham and Women's Hospital; and <sup>§</sup>Department of Pediatrics, Children's Hospital Medical Center, Harvard Medical School, Boston, Massachusetts 02115*

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M.H. Sayegh and N.L. Tilney are co-senior authors.

Address correspondence to Dr. Mohamed H. Sayegh, Brigham and Women's Hospital, 75 Francis St., Boston, MA 02115. Phone: 617-732-5259; FAX: 617-732-5254; E-mail: sayegh@bustoff.bwh.harvard.edu

*Received for publication 13 March 1997 and accepted in revised form 28 May 1997.*

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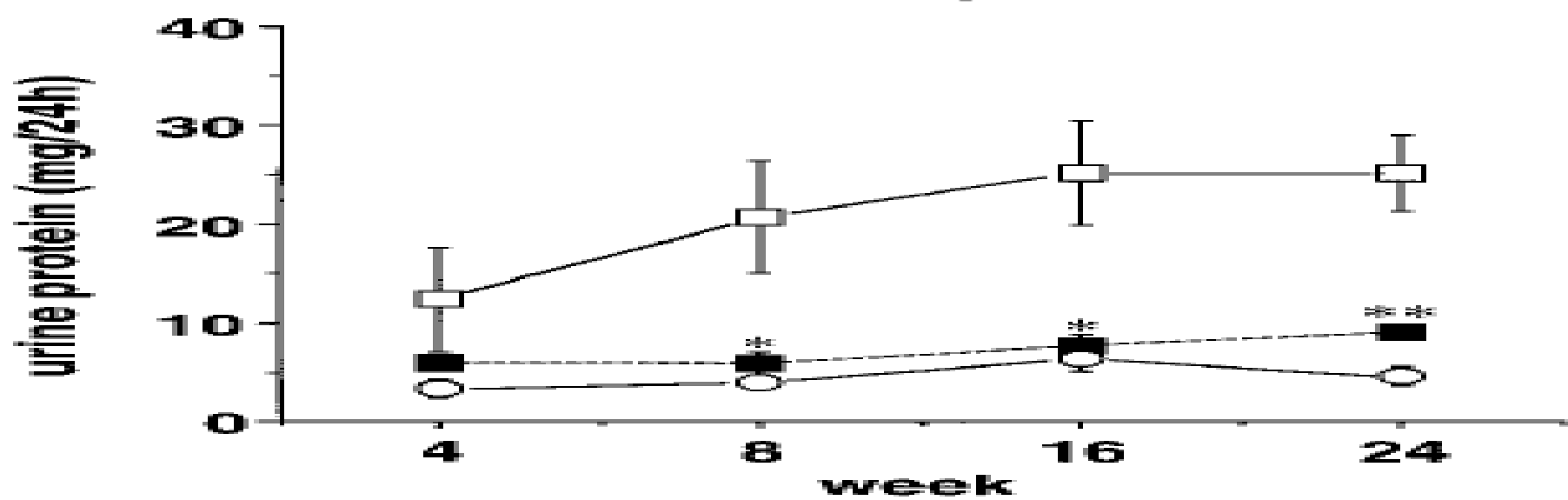
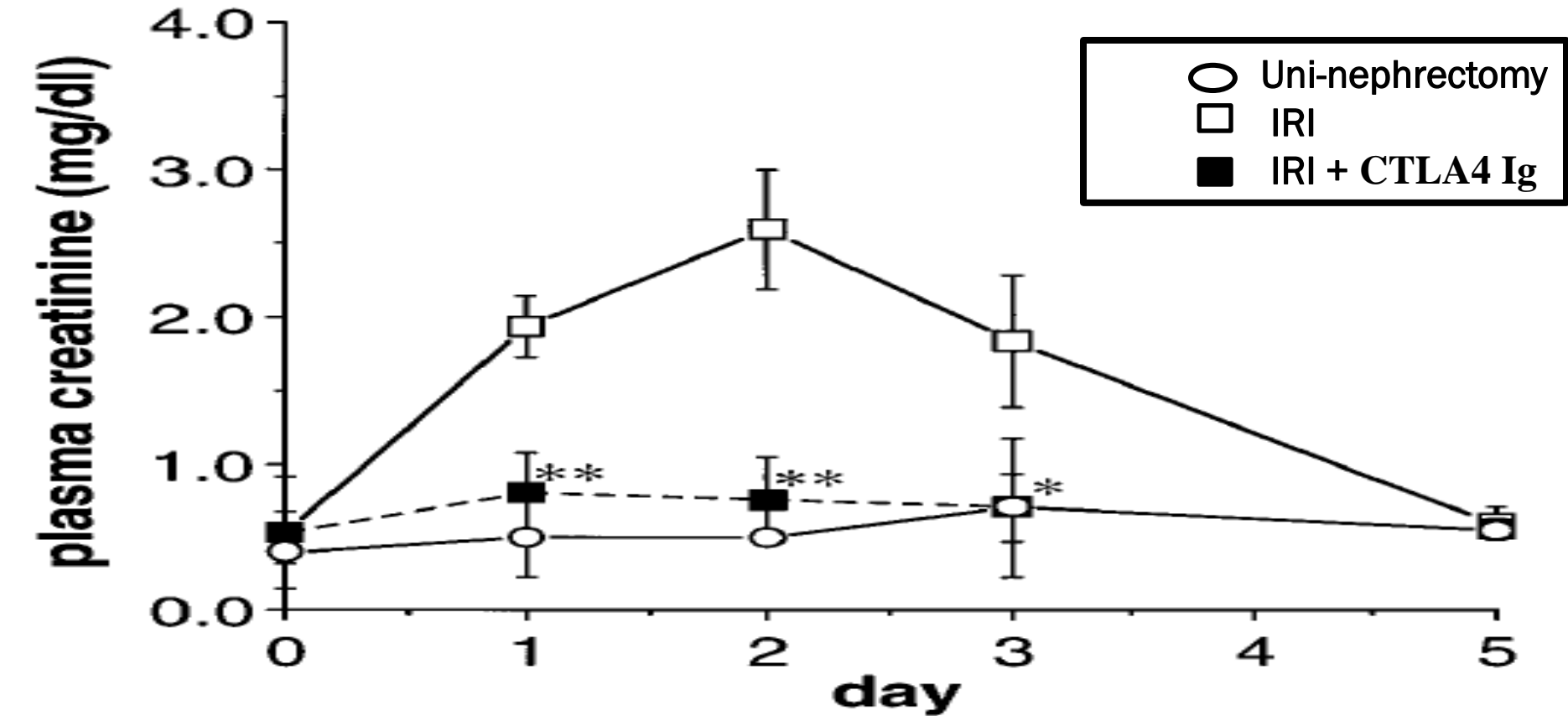
J. Clin. Invest.

© The American Society for Clinical Investigation, Inc.  
0021-9738/97/09/1199/05 \$2.00

Volume 100, Number 5, September 1997, 1199–1203

<http://www.jci.org>

**Blockade of T cell CD28-B7 costimulation with CTLA4Ig Ig both on the day of renal IRI and during the first week after IRI in uninephrectomized rats**





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# T OR B LYMPHOCYTES IN RENAL INJURY

## ??????????????



## Phenotypic and Functional Characterization of Kidney-Infiltrating Lymphocytes in Renal Ischemia Reperfusion Injury

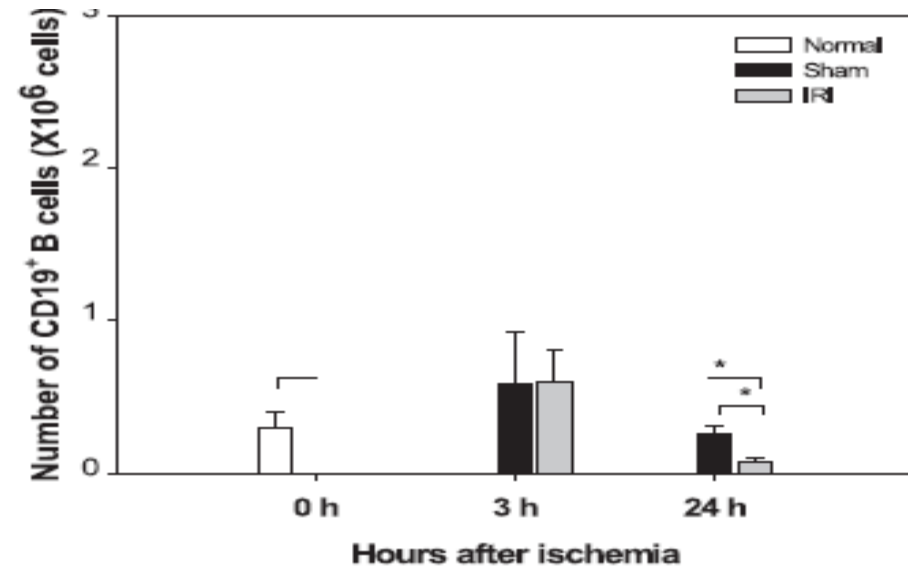
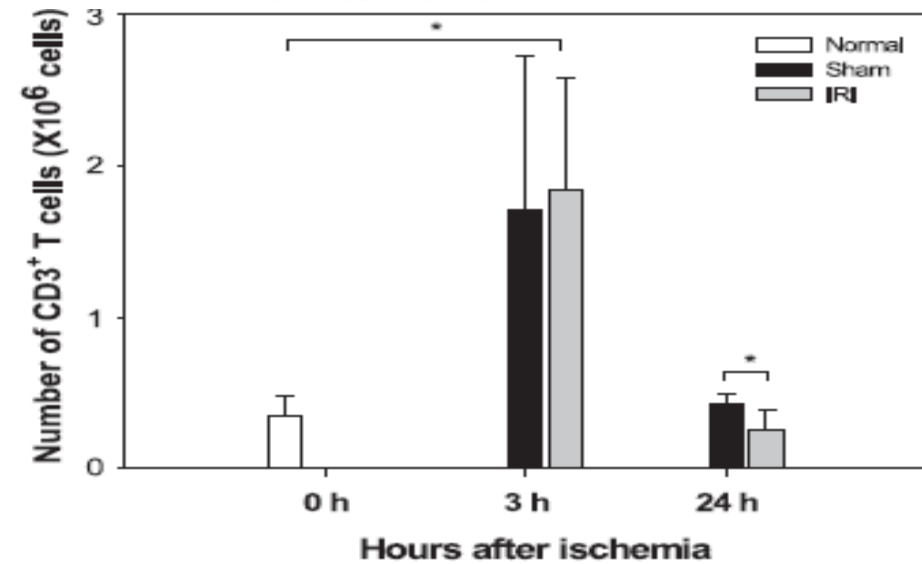
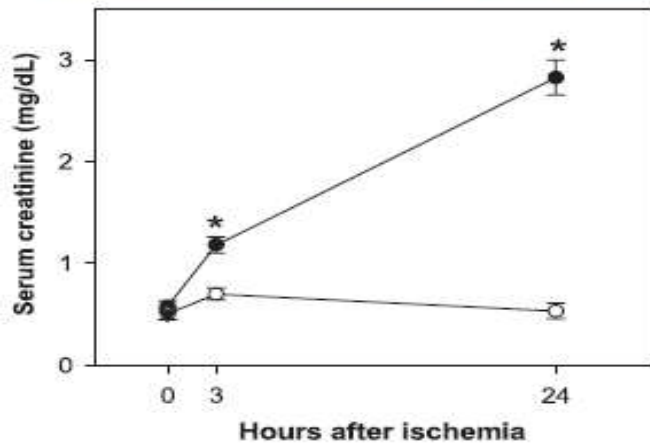
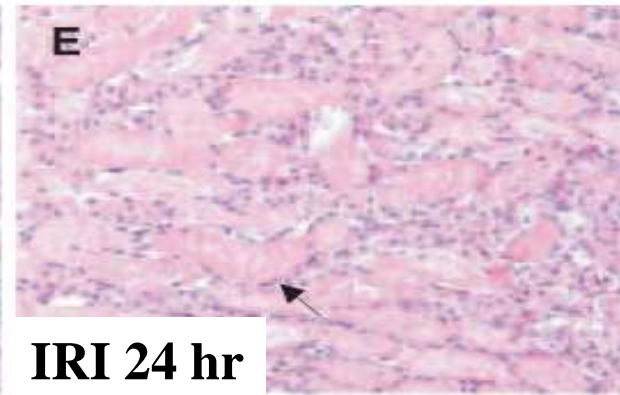
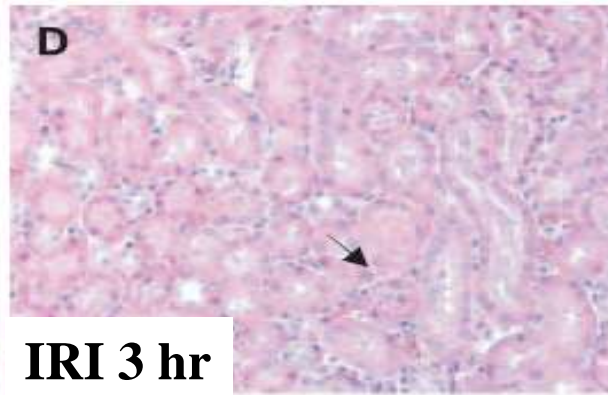
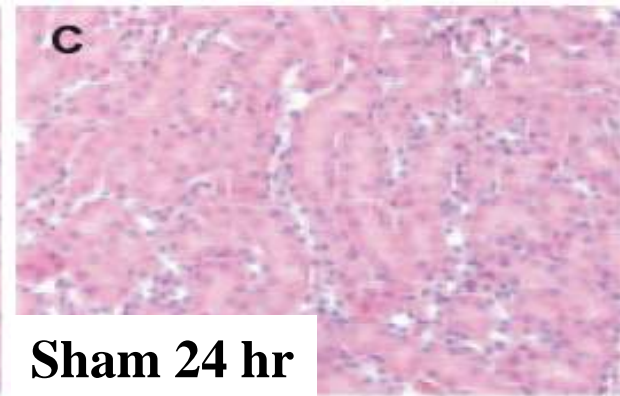
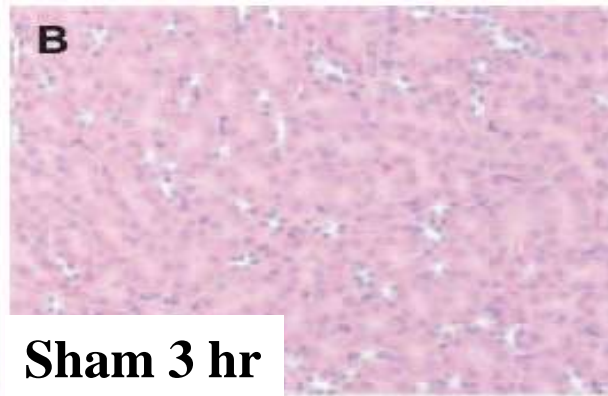
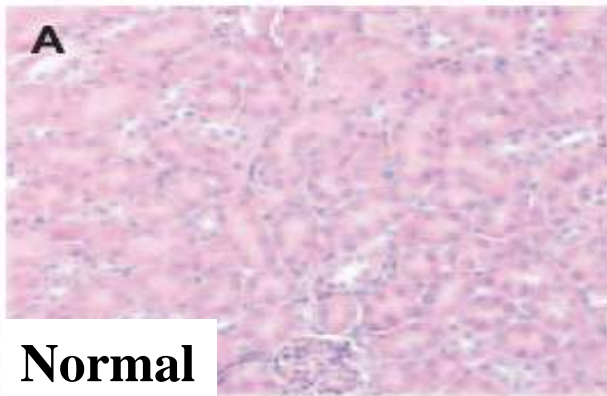
This information is current as of November 25, 2012.

Dolores B. Ascon, Sergio Lopez-Briones, Manchang Liu, Miguel Ascon, Vladimir Savransky, Robert B. Colvin, Mark J. Soloski and Hamid Rabb

*J Immunol* 2006; 177:3380-3387; ;  
<http://www.jimmunol.org/content/177/5/3380>

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**Male C57BL/6J wild-type were bluntly dissected and a microvascular clamp was placed on each renal pedicle for 30 min and compared with Sham operated animals.**





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**CD4 OR CD8 T LYMPHOCYTES**

**??????????**

# Identification of the CD4<sup>+</sup> T cell as a major pathogenic factor in ischemic acute renal failure

Melissa J. Burne,<sup>1</sup> Frank Daniels,<sup>1</sup> Asmaa El Ghandour,<sup>1</sup> Shamila Mauiyyedi,<sup>2</sup>  
Robert B. Colvin,<sup>2</sup> Michael P. O'Donnell,<sup>1</sup> and Hamid Rabb<sup>1</sup>

<sup>1</sup>Division of Nephrology, Hennepin County Medical Center, University of Minnesota, Minneapolis, Minnesota, USA

<sup>2</sup>Department of Pathology, Massachusetts General Hospital, Harvard Medical School, Boston, Massachusetts, USA

Address correspondence to: Hamid Rabb, Johns Hopkins School of Medicine, Ross Building, Room 970, 720 Rutland Avenue, Baltimore, Maryland 21205, USA. Phone: (410) 502-1555; Fax: (410) 614-5129; E-mail: hrabbl@jhmi.edu.

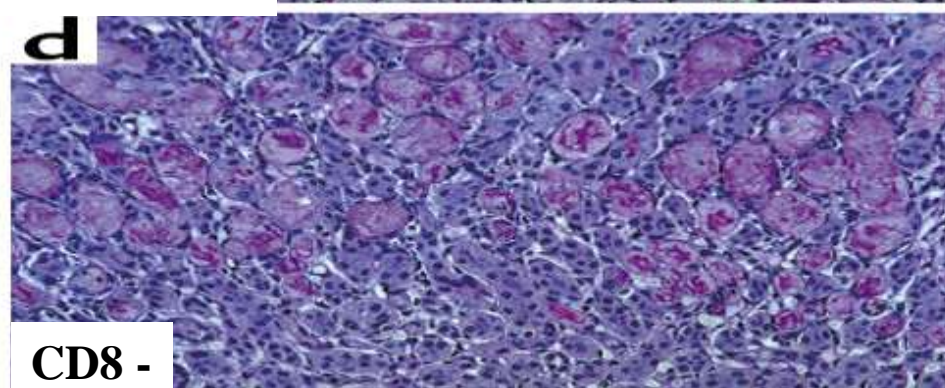
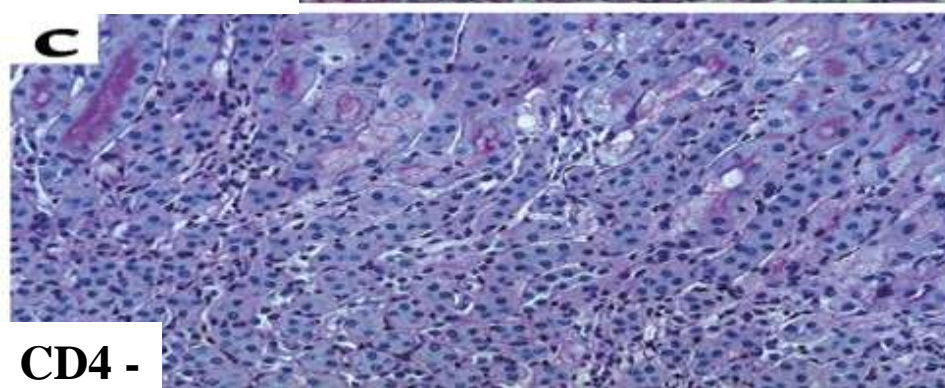
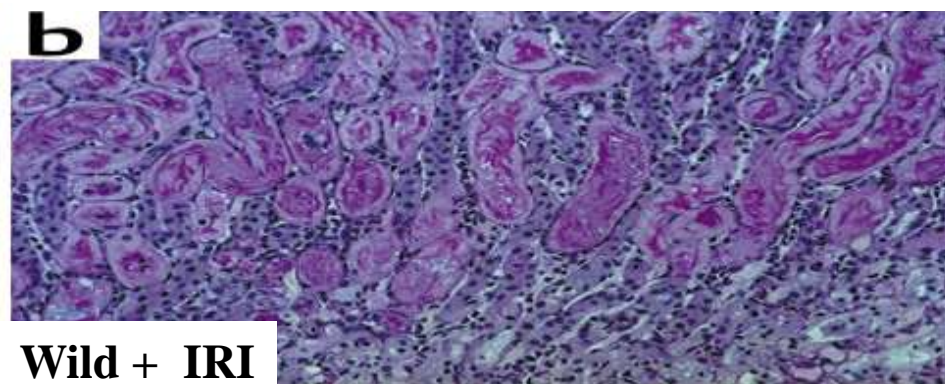
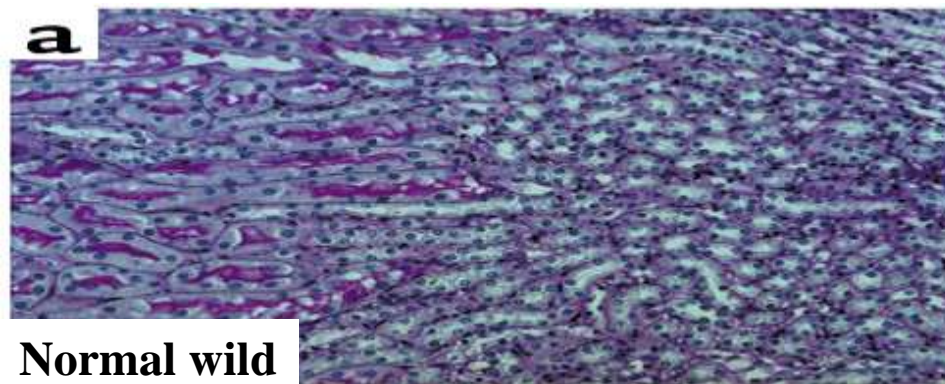
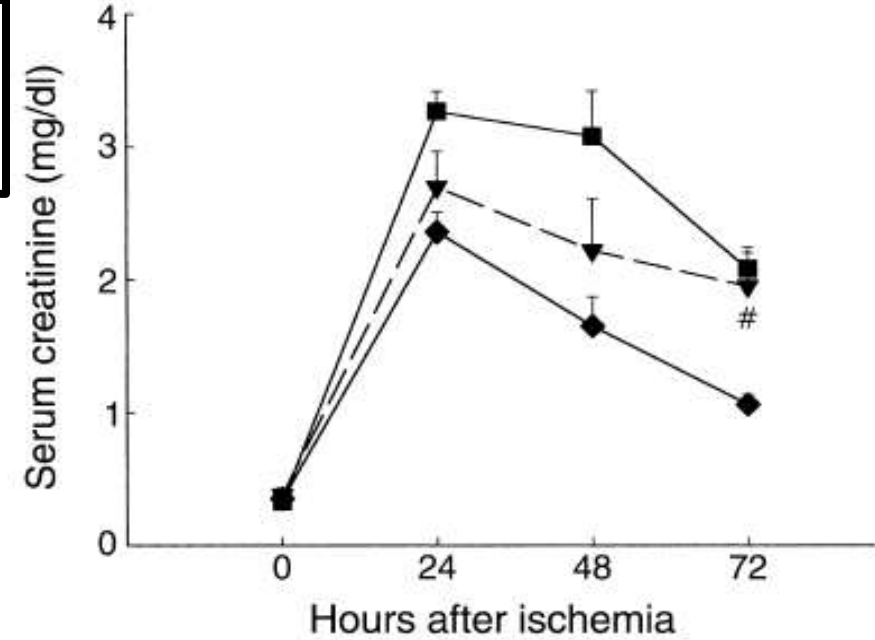
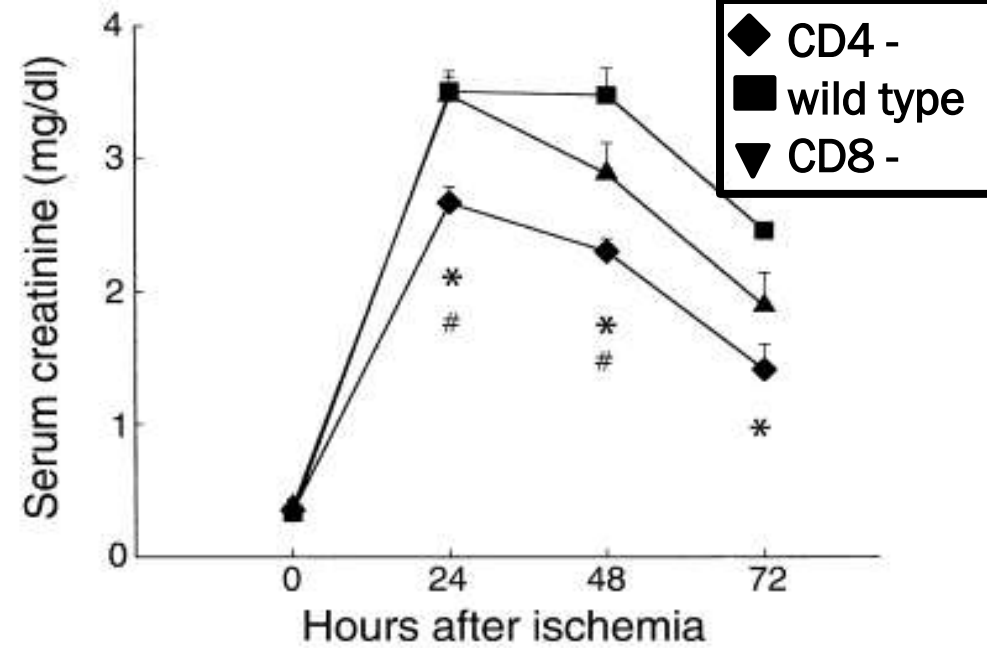
Received for publication December 27, 2000, and accepted in revised form September 17, 2001.

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The Journal of Clinical Investigation | November 2001 | Volume 108 | Number 9

**They compared IRI in C57BL/6 wild-type, *nu/nu* mice & *nu/nu* mice were adoptively transferred with wild-type T cells.**







- 
- ✗ CD4 T-cell has 2 subsets:
    - Th1 phenotype: pathogenic
    - Th2 phenotype: protective

---

**DOES LYMPHOCYTES HAVE ANY ROLE IN  
RENAL REPAIR ????????**

# Regulatory T Cells Suppress Innate Immunity in Kidney Ischemia-Reperfusion Injury

Gilbert R. Kinsey, Rahul Sharma, Liping Huang, Li Li, Amy L. Vergis, Hong Ye, Shyr-Te Ju, and Mark D. Okusa

Division of Nephrology and Center for Immunity, Inflammation and Regenerative Medicine, University of Virginia Health System, Charlottesville, Virginia

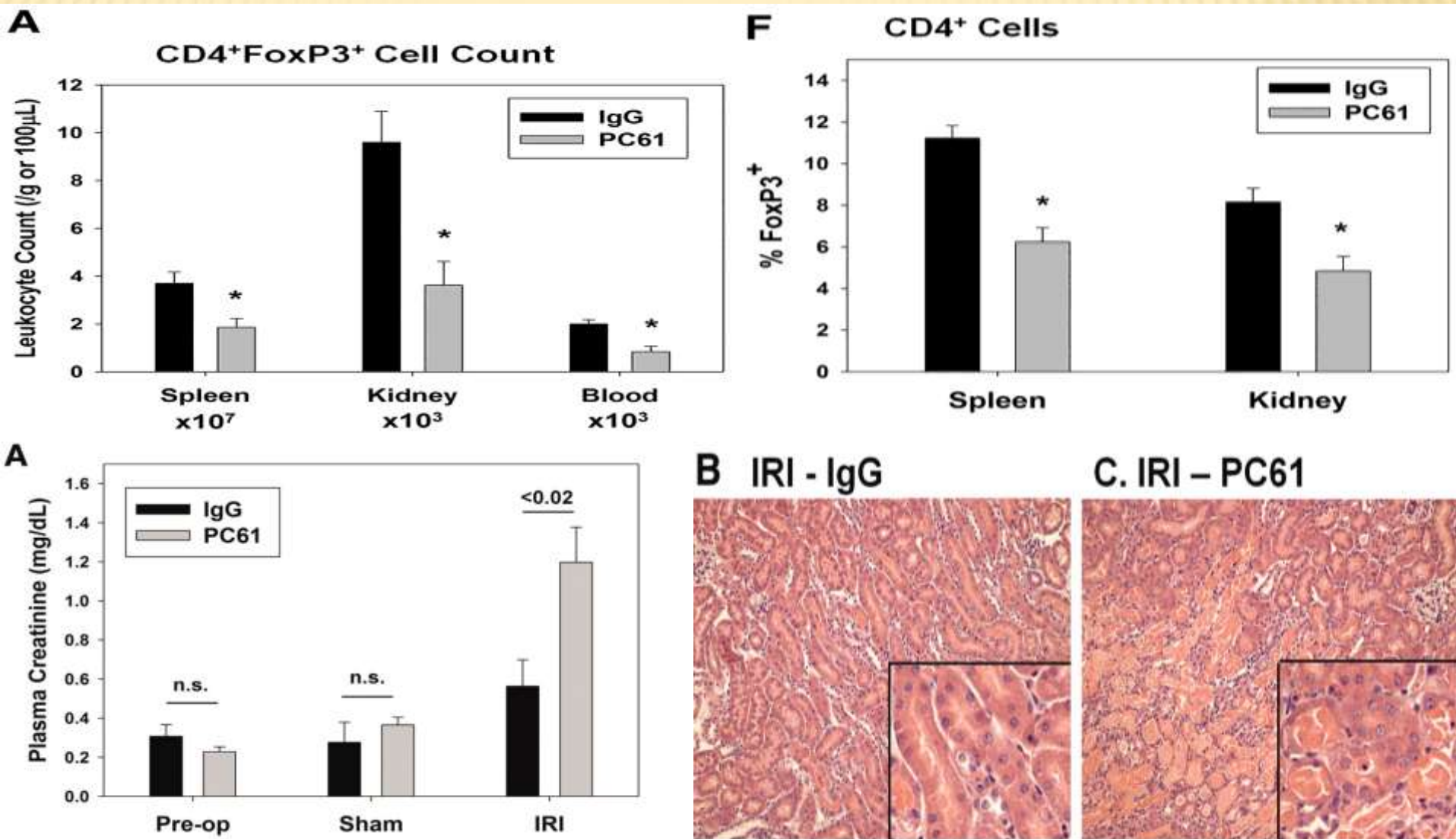
*J Am Soc Nephrol* 20: 1744–1753, 2009. doi: 10.1681/ASN.2008111160

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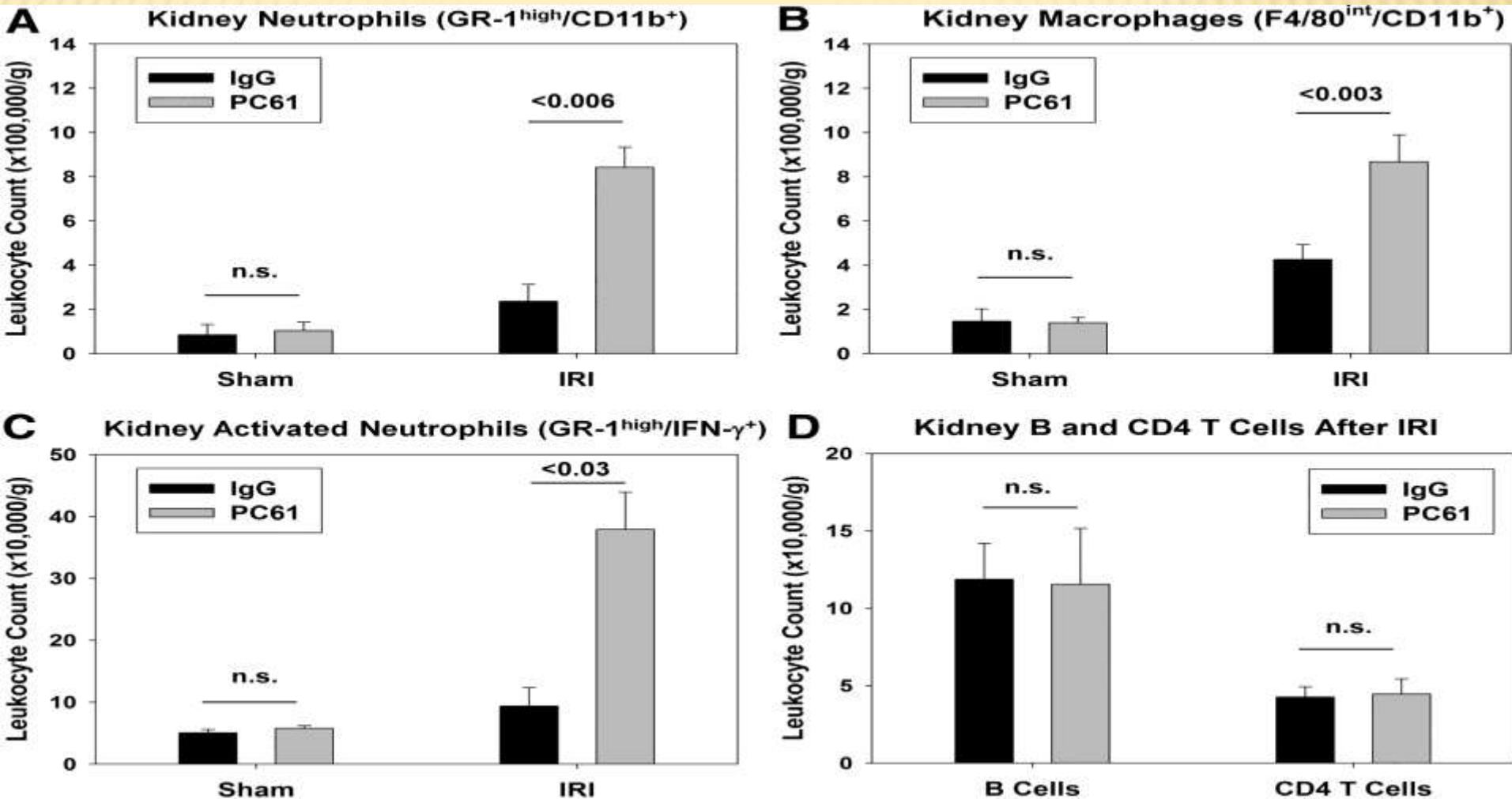
**Partial depletion of Tregs with an anti-CD25 mAb before IRI. Adoptive transfer of lymph node cells from wild-type mice or FoxP3-deficient Scurfy mice into T cell– and B cell– deficient RAG-1 knockout mice to generate mice with and without FoxP3 Tregs, respectively.**



# CD25 MAB (PC61) ADMINISTRATION REDUCES Tregs & POTENTIATES IRI

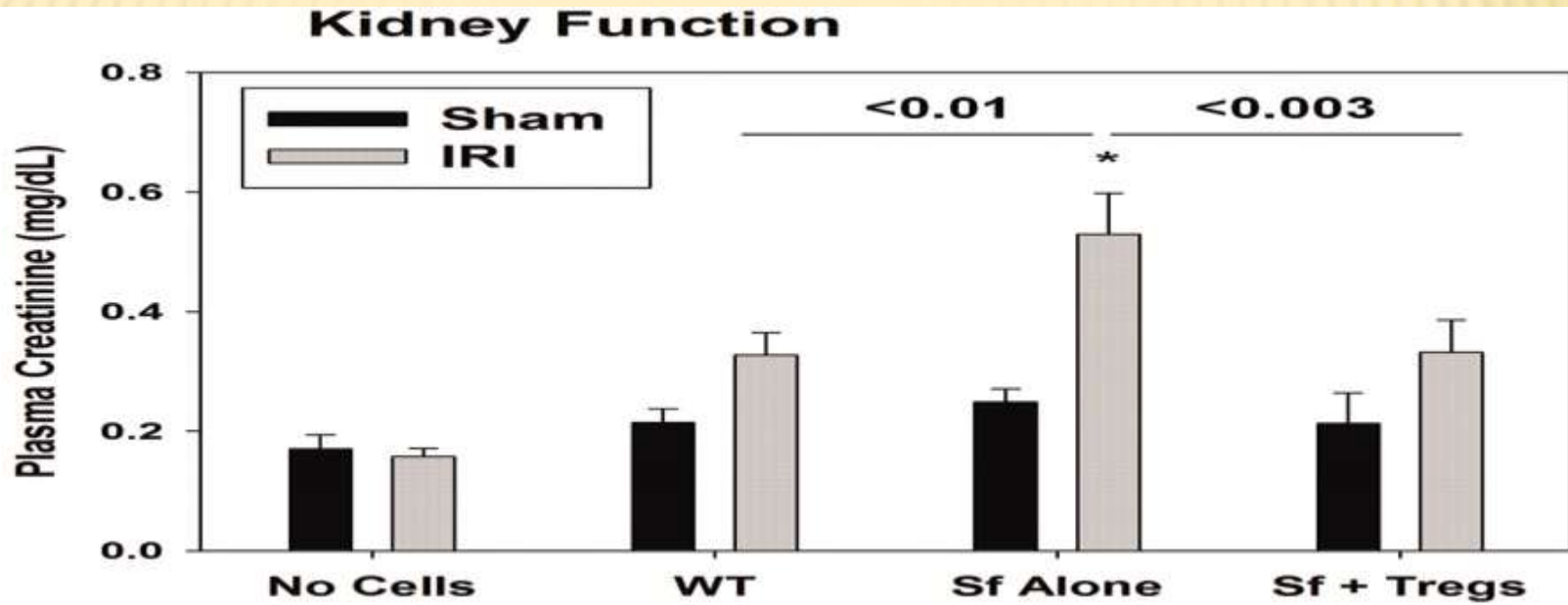


# PARTIAL Treg DEPLETION ENHANCES LEUKOCYTE ACCUMULATION AFTER IR.

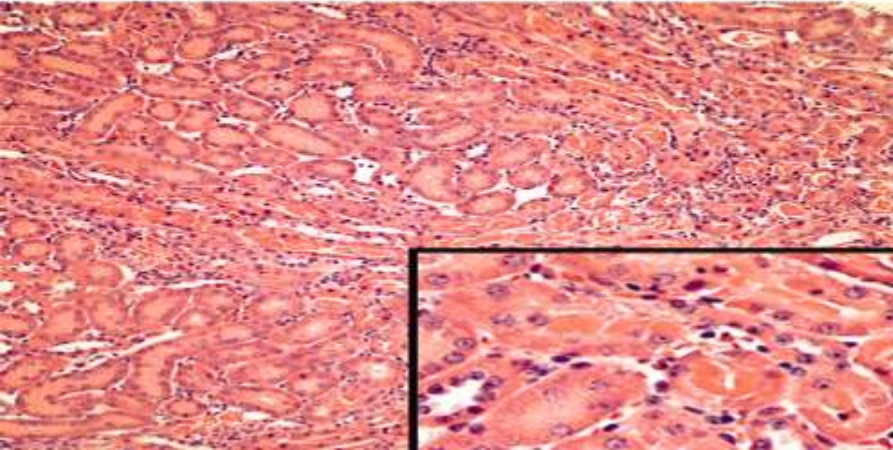


# Treg REPLETION INHIBITS KIDNEY IRI

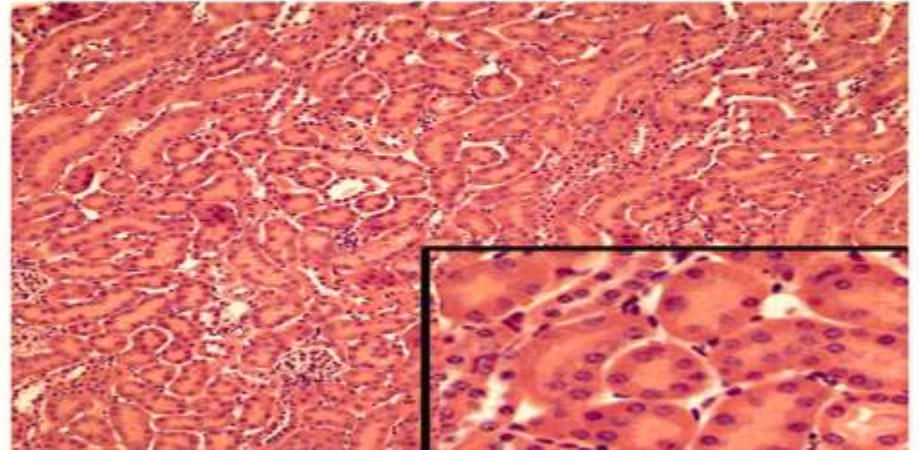
**A**



**B Sf → RAG-1 KO - IRI**



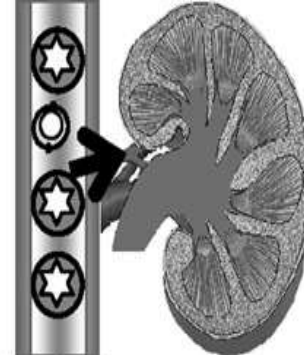
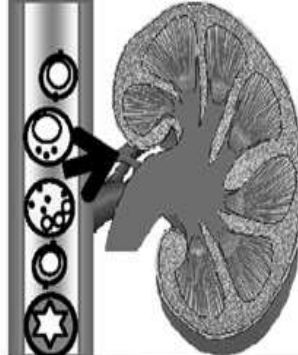
**C Sf + Tregs → RAG-1 KO - IRI**







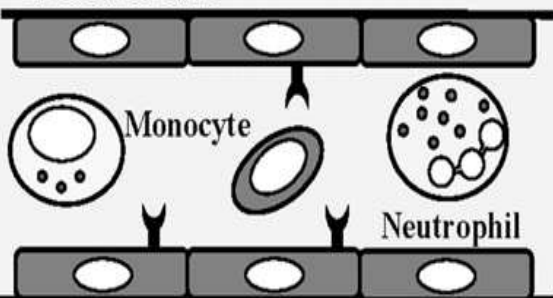
Ischemia-reperfusion



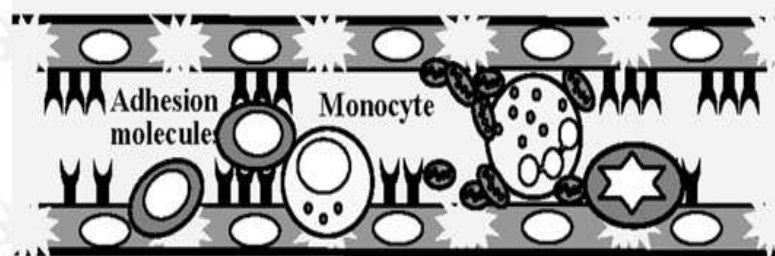
Normal kidney

Post-ischemic kidney

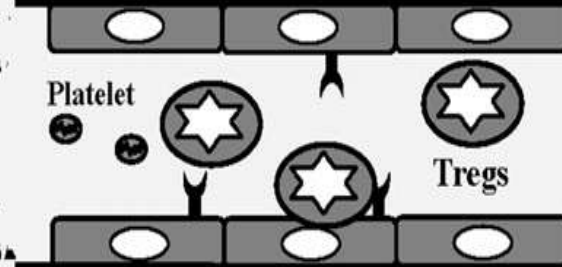
Endothelium



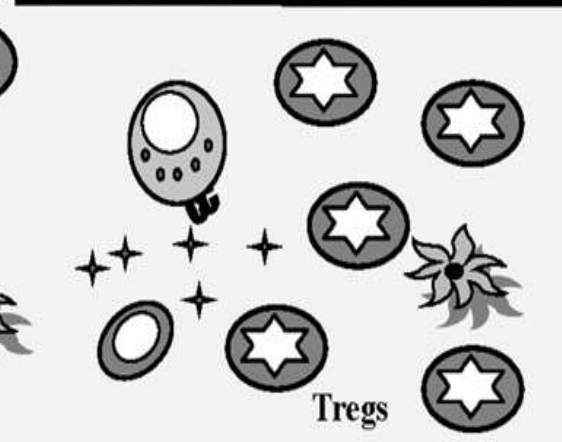
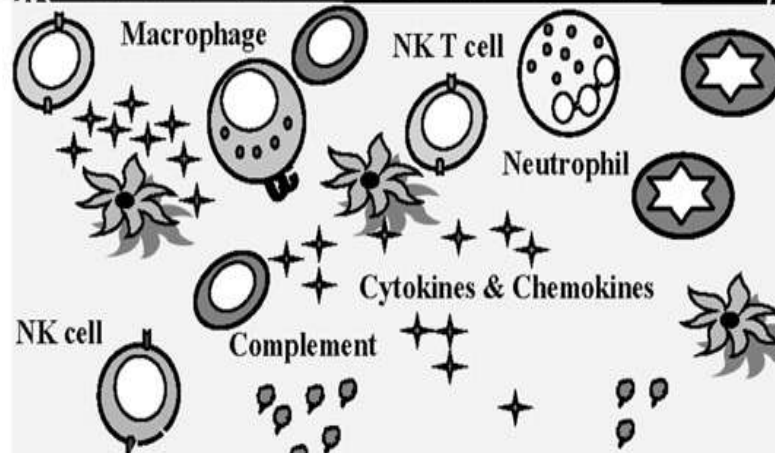
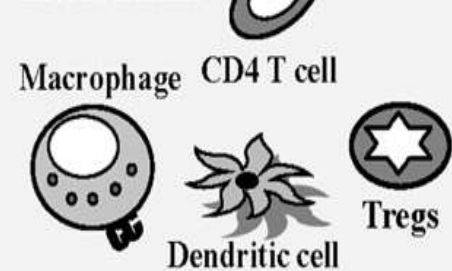
Early injury phase



Repair phase



Interstitial



Tubular epithelium



# CONCLUSION

- ✖ Macrophages and distinct T-cell subsets are capable of anti-inflammatory as well as proinflammatory functions.
- ✖ Wound-healing macrophages (M2a) , regulatory macrophages (M2c) and regulatory T cells (Tregs) are remarkably potent at reducing renal injury in a number of different renal disease models.
- ✖ Understanding the biology of macrophages and lymphocytes and their ability to repair renal tissue will enable the future discovery of therapies for kidney diseases.

*Thank you*

